

Quality Characteristics of Soy-akamu Powder Formulated from Sorghum and Sprouted Soybean Flour Blends for Complementary Feeding

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Abstract: Soy-akamu is nutritionally poor hence the improvement by formulating with sprouted soybean to fortify and restore protein and other nutrients lost during processing to prevent hidden hunger. Sprouted soybean obtained from hand dehulled 12h tap water steeped sorted soybean, 72h sprouted and 20min boiled in 0.05% sodium bicarbonate solution was milled with 72h steeped, drained and washed cleaned sorghum seeds according to 75: 25, 50: 50 and 25: 75 sorghum; sprouted soybean blends. Blended pastes were sieved and dewatered separately with double layered calico cloth to obtain the pastes. Proximate results showed increase in dried matter content (85.94 to 86.68%) with increase in soybean paste inclusion, moisture content decreased (14.06 to 13.32%) protein increased (5.11 to 39.96%), fat increased (1.51 to 11.21%), fiber increased (1.41 to 4.82%), ash increased (0.46 to 4.61%), carbohydrate decreased (77.44 to 26.07%) and energy increased (335.26 to 364.99Kcal). Bulk density increased (0.33% to 0.66g/ml), viscosity decreased (117.02 to 84μPas), swelling power increased (22.83 to 30.04), gelatinization temperature decreased (66.00 to 45°C) and gelatinization time increased (0.35 to 0.37sec). Gruel from 100% sorghum scored the highest acceptability. Sprouted soybean blending showed an improvement in the nutrients content of soy-akamu and decrease in acceptability beyond 25% inclusion.

Keywords: Proximate Composition, Functional Properties, Sensory Property, Akamu Paste, Complementary Feeding

1. Introduction

Akamu in Igbo or *ogi* in Yoruba is a fermented cereal paste made from maize (*Zea mays*), sorghum or guinea corn (*Sorghum vulgare*) [1] or millet (*Pennisetum tyopideum*). The colour of akamu depends on the cereal colour used which may be cream colour from maize, reddish-brown colour from sorghum [2, 1]. Akamu from sorghum is popular in northern Nigeria and most parts of West Africa which porridge or gruel is very smooth in texture and has a sour taste (pH 4.8) and inhibits growth of some bacteria [3]. Fortification of *ogi* with legumes, vitamins and minerals alongside improvement on the production techniques which led to the development of *soya-ogi*, a combination of maize and soybeans have been reported [4].

Increase food use of sorghum in sub-Saharan Africa will

alleviate the problem of chronic under-nourishment, as sorghum is readily available in all parts of the countries [2, 1]. However, nutritional problems associated with cereals are their general low protein content and significant less digestible nature when cooked compared to other cooked cereal protein [1]. This could cause protein malnutrition for infants fed exclusively on sorghum based meal. Also, cereals are low in minerals and vitamins which along with protein are essential nutrients needed for the wellbeing and healthy growth and development of infants, children and adults who relish exclusively on akamu. Supplementation of cereals with locally available legumes that is high in protein will increase protein quality and the nutritive values of akamu lost during steeping, milling and sieving processes compared to the use of single cereal [5, 6]. Also, protein supplementation will

prevent protein-energy malnutrition in infants weaned exclusively on *akamu*. As cereals and their products are invariably cooked prior to consumption, these problems are inevitable and require urgent attention.

Several efforts are currently on, in Africa to modify the processing of *akamu* with a view to enhancing its nutritive value, shelf life and possible therapeutic qualities [2, 7]. One likely method of achieving this is by formulation with soybeans alone [8] or by formulation or may be fortified with vitamins and minerals [4]. Also, cereal fermentation was employed to preserve, impart aroma and flavour while producing novelty foods products with improved nutrients for complimentary feeding [9, 10].

Soybean oil varies between 13.9 to 23.2% depending on locality, cultivar and horticultural practices [11] and 40% higher calorific value than protein and carbohydrates [12]. Soybean is a cheap source of good quality protein with good balance of the essential amino acid and high quality oil [13]. Protein content of soybean contains considerable quantity of lysine (6.2g/16gN), but limited by methionine and cystine content (2.9g/16gN) [14]. Absence of cholesterol, lactose and presence of essential amino acids makes soybean vital for infant growth and maintenance.

Soybean sprouting is an age long act of improving the nutritive value of legumes, reduction in anti-nutrients and flatulence causing oligosaccharides (stachyose and raffinose), thereby increasing protein digestibility and sensory properties [15]. Also, sprouting increases vitamins, minerals (calcium, copper, manganese, and zinc), nutrient bioavailability and free amino acid. Sprouted soybean had been used to formulate complementary foods [16].

Gruel also known as *akamu* or *ogi* when prepared from the paste with hot water after dissolving same with tap water into light thick consistency serves as a major weaning food for the infants in West Africa [17] or by nursing mothers as it encourages breast milk supply, a good vehicle for food nutrients to sick people because it is an energy food rich in carbohydrates with traces of vitamins (Oyelana *et al*; 2012). it is easily digested and light in the stomach [18]. A blend of sorghum- sprouted soybean will not only boost the nutritive value of *akamu* as a breakfast meal, weaning food and choice food for the sick [17] but will also boost production and consumption of soybean [19]. This work therefore aimed at improving the protein content of *akamu* using sprouted soybean paste.

2. Materials and Methods

2.1. Material Procurement

The sorghum and soybean used for this work were purchased from Urbani, Ibeku ultra modern market Umuahia Abia State, Nigeria.

2.2. Preparation of Sorghum-soybean Paste (*Akamu*)

Sorghum was steeped with clean tap water for 72h, decanted, washed and wet milled (Power Delux Electric

Blender Model PDB-8231-F) together with the sprouted soybean cotyledons according to formulation (Table 1). Sprouted soybean cotyledons were obtained by sorting/cleaning, steeping (12 hours), sprouting (72h), washing, boiling with 0.05% sodium bicarbonate for 20min, and hand dehulled. The paste was then sieved with calico cloth, allowed to sediment, and dewatered with a double layered calico cloth to obtain sorghum-soybean wet paste or *akamu* (Figure 1).

Steeped sorghum and sprouted dehulled soybean cotyledons were dried in oven at 60°C to constant weight, milled and sieved separately after cooling, and blended (Table 1) to obtain flour blends [20].

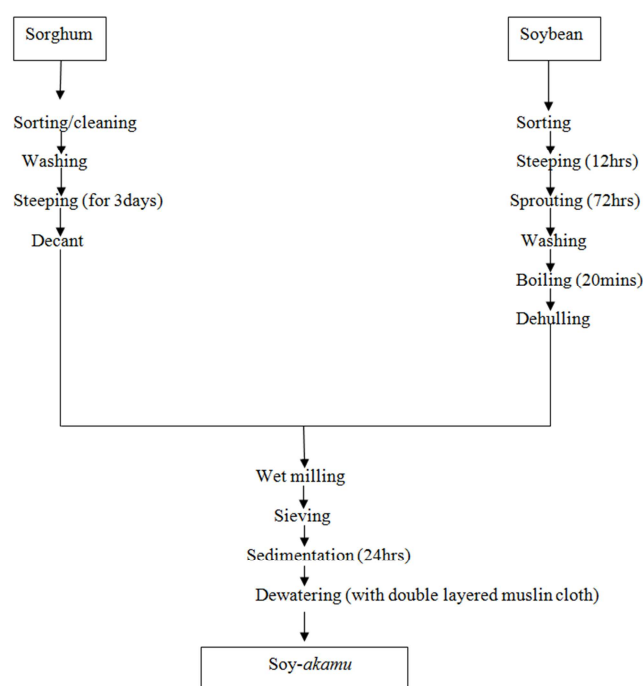


Figure 1. Flow chart for production of sorghum-sprouted soybean *akamu* paste.

Table 1. Formulation blends for sorghum and sprouted soybean.

Sample	Sorghum (%)	Sprouted soybean
101	100	0
104	75	25
106	50	50
108	25	75

2.3. Proximate Analyses

2.3.1. Moisture Content

Moisture content was determined according to AOAC [21] protocol. Five grams (5g) of each sample were placed into a previously washed, dried and weighed moisture cans, dried in the oven at 105°C for 3 and placed in a desiccator to cool. Thereafter the process was repeated at a chosen time interval other than 3hr until a constant weight was obtained. The weight of the moisture lost was calculated as a percentage of weight of sample analyzed as expressed below.

$$\% \text{ Moisture content} = \frac{W_2 - W_3}{W_2 - W_1} \times \frac{100}{1}$$

Where W_1 =weight of the empty moisture can, W_2 =weight of moisture can + sample before drying and W_3 =Weight of moisture can + sample dried to constant weight.

2.3.2. Crude Fiber Content

Five grams (5g) of the sample was boiled for 30min with 150ml of a solution containing 1.25g H_2SO_4 per 100ml under reflux. The solution was filtered through a two-fold muslin cloth on a fluted funnel and washed with boiling water until it is no longer acidic. The residue was returned to the flask and boiled for 30min with 150ml of solution containing 1.25g of carbonate free NaOH per 100ml. After washing, the sample was allowed to drain dry before being transferred in a weighed crucible where it was dried in the oven at $105^\circ C$ to a constant weight which was recorded. The sample was finally incinerated in a muffle furnace and the weight of the ash was taken which was used to determine the fiber thus:

$$\% \text{ Crude fiber} = 100 (W_2 - W_3)$$

Where: W_2 =Weight of crucible + sample after oven drying and W_3 =Weight of crucible + sample after incineration [22].

2.3.3. Fat Content

Soxhlet fat extraction method [22] was used. The boiling flask were dried in the oven and cooled in the dessicator. Two gram (2g) of each sample was placed in the marked thimbles and weighed. The boiling flask was filled with 300ml of petroleum ether at $60^\circ C$. The Soxhlet apparatus was assembled and allowed to reflux for 5-6h thereafter the thimble was removed, petroleum ether collected and the flask dried at $105^\circ C$ for 1hour. It was cooled and weighed. Fat content was calculated as follows:

$$\% \text{ oil} = W_2 - W_1 / W_3 \times 100/1$$

Where: W_1 =Weight of empty flask, W_2 =Weight of flask + extract and W_3 =Weight of sample used.

2.3.4. Protein

The Kjeldahl method [23] was used. Two grams (2g) of the samples were mixed with 10ml of concentrated tetraoxosulphate (VI) acid in a Kjeldahl digestion flask. A tablet of selenium catalysts was added and the mixture was heated inside a fume cupboard. The digest was transferred into a 100ml volumetric flask, made up with equal volume of 45% sodium hydroxide (NaOH) solution and poured into a Kjeldahl distilled apparatus. The mixture was distilled and the distillate was collected into a 4% boric acid solution containing 3 drop of zuazaga indicator (mixture of methyl red and bromacresol green). A total of 15ml distillate was titrated against 0.02N tetraoxosulphate (VI) acid solution until the colour changed from green to a deep red or pink end point. Total nitrogen was calculated and multiplied with the factor 6.25 to get the crude protein content.

$$\% \text{ Crude protein} = \% N \times 6.25$$

$$\% N_2 = (100 \times N \times 14 \times V_F / W \times 1000 \times V_A) \times t/1$$

Where: W =Weight of the sample, N =Normality of the filtrate, V_F =Total volume of the digest=100ml, V_A =Volume of the digest distilled and T =Titer volume.

2.3.5. Carbohydrate

Carbohydrate content of the paste samples was calculated by difference [23] thus:

$$\% \text{ CHO} = 100\% - \% (a + b + c + d + e)$$

Where: CHO=carbohydrate, a=protein, b=fat, c=crude fiber, d=ash and e=moisture.

2.3.6. Ash Content

Ash content was determined by furnace incineration gravimetric method [22]. Five grams (5g) of the sample were placed into washed, oven dried and weighed porcelain crucible. The sample was incinerated at $550^\circ C$ for 3h to grayish ash in a muffle furnace, cooled in a desiccator and weighed. Weight of the ash obtained was determined b/y difference and expressed as a percentage of the weight of the samples incinerated as shown:

$$\% \text{ ash} = 100 \times [W_3 - W_1] / [W_2 - W_1]$$

Where:

W_1 =Weight of empty crucible, W_2 =weight of crucible +food before drying and/or ashing and W_3 =Weight of crucible +ash.

2.3.7. Energy Values

Proximate values of crude fat, protein, fiber and carbohydrate (Table 1) were used for the calculation of energy levels of the samples using Atwater factor [24].

2.4. Functional Properties

Functional properties of sorghum and soybean flour blends were determined.

2.4.1. Bulk Density

Ten grams (10g) of the sample were measured into 100ml graduated measuring cylinder and tapped on repeatedly on a pad placed on a laboratory bench until no further reduction in volume occurred [22]. Bulk density was determined as the ratio of the weight of the sample to its volume as shown below:

$$\text{Bulk density} = W/V \text{ (g/dm}^3\text{)}$$

Where: W =weight of sample in grams and V =volume of sample (dm^3)

2.4.2. Viscosity

Ten grams (10g) flour was suspended in 100ml distilled water and mechanically stirred for 2h at room temperature and the viscosity was measured using bench rotary viscometer, NDS-55 Surgifriend Medical England [22].

2.4.3. Gelatinization Temperature

Ten grams (10g) of the flour sample were suspended in 100ml of water in a test tube. The aqueous suspension was heated in a boiling water bath with continuous stirring and the temperature at which gelatinization was visually noticed was recorded as the gelatinization temperature [22].

2.4.4. Swelling Power Index

Swelling power is a measure of the hydration capacity of starch expressed as the weight of centrifuged swollen granules, divided by the weight of the original dry starch used to make the paste. Two grams (2g) of the flour samples were suspended in 10ml of water and incubated in a thermostatically controlled water bath at 95°C in a tarred screw cap tube of 15ml. The suspension was stirred intermittently for 30min period to keep the starch granules suspended. The samples were then rapidly cool to 20°C with cool water and centrifuged/ at 2200× g for 15 minutes to separate the jelly and the supernatant [25].

$$\text{Swelling power} = W_2 \times 100 / W_{dm} \text{ (100-solubility)}$$

Where: W_2 =W/eight of centrifuged swollen granules and W_{dm} =weight of original dry sample

2.4.5. Gelatinization Capacity

The method of Onwuka [22] was used. Five grams (5g) of each flour sample was dissolved in 100ml of distilled water in a 500ml beaker to obtain 5% sample suspension. Two (2ml) of the sample was dissolved in 5ml distilled water in attest tube, heated for 1hr in a boiling water bath and cooled rapidly under running cool tap water. The test tube was cool further for 2h at 4°C and inverted. The gelation capacity is the least gelation concentration determined when the sample from inverted test tube will not fall o/r slip.

3. Sensory Evaluation

Twenty (20) semi trained panelists randomly selected from the students of Michael Okpara University of Agriculture, Umudike within the age bracket of 18 to 30yr who are familiar with the gruel were used. The coded/ samples were

presented to each panelist separately in same types of saucers along with bottled water /under bright illumination. They were to taste the sample one after the other, rinse their mouths after each tasting and score the samples using 9-point Hedonic scale [27] where 9 represents dislike extremely, 1 dislike extremely, and 5 neither like nor dislike. Appearance, flavour, taste, texture and general acceptability were evaluated [26].

4. Statistical Analyses

Data obtained from the analyses were subjected to analysis of variance (ANOVA) using SPSS software package version 20. Means were separated using Duncan Multiple Range test to determine the significant difference at 5% probability.

5. Results and Discussion

5.1. Proximate Composition

Table 1 summarized the proximate composition results of the sorghum - sprouted soybean paste blends.

5.1.1. Dry Matter

Percentage dry matter (DM) content of the paste samples ranged from 85.78% in sample 104 (100% sorghum) to 86.68% in 108 (25% sorghum: 75% sprouted soybean). Significant difference ($p < 0.05$) between all the dry matter content of the samples prefigured significant variations in dry matter contributions of various blends in the formulations. Lower dry matter content of sample 104 (100% sorghum) than the rest samples indicated that sorghum had more moisture than the rest blends most especially 25% sorghum and 75% sprouted soybean probably due to higher proportion of sprouted soybean. Soybean is a legume which protein digestibility and free amino acids increases with sprouting [15, 28]. Proteins bound and hold water [29] which may have contributed in lowering sample moisture content as substantiated in the table which showed dry matter increase with increase in soybean inclusion.

Table 2. Proximate Composition of ogi Paste from Sorghum and Sprouted soybean Blends (%).

Sps DM/	MC	CP	EE	CF	ASH	CHO	EV (Kcal)
10185.94/ ^f ±0.03/	14.06 ^b ±0.03	5.11 ^g ±0.02	1.51 ^g ±0.00	1.41 ^g ±0.01	0.46 ^d ±0.00	77.44 ^b ±0.00	335.26 ^d ±0.06
10/485.78 ^g ±0.01/	14.22 ^a ±0.01	17.20 ^g ±0.00	6.55 ^c ±0.00	2.81 ^g ±0.01	1.77 ^c ±0.02	57.43 ^d ±0.00	348.93 ^b ±0.04
10/685.84 ^g ±0.01/	14.16 ^a ±0.01	25.68 ^c ±0.03	9.85 ^c ±0.00	3.94 ^c ±0.01	2.85 ^b ±0.01	43.52 ^f ±0.01	364.99 ^a ±0.01
10/886.68 ^c ±0.03/	13.32 ^c ±0.03	39.96 ^a ±0.01	11.21 ^a ±0.02	4.82 ^a ±0.01	4.61 ^a ±0.01	26.07 ^b ±0.00	335.70 ^c ±0.14

Values are mean triplicate determinations±standard deviation. Values with the same superscript within the same column are not significantly different ($p < 0.05$) while those with different superscript are significantly different. Sps=samples, DM=Dry matter, MC=Moisture contents, CP=Crude protein, EE=Ether extract, CF=Crude fiber, EV=Energy value, CHO=Carbohydrate. 101=100% sorghum 104=75% sorghum and 25% sprouted soybean, 106=50% sorghum and 50% sprouted soybean and 108=25% sorghum and 75% sprouted soybean.

5.1.2. Moisture Content

Increase in moisture contents (MC) of the paste samples from 13.32% in sample 108 (25% sorghum: 75% sprouted soybean) to 14.22% in sample 104 (75% sorghum: 25% sprouted soybean) validated the inverse relationship of DM

with moisture earlier reported. The difference could be traced in part to the variations in proportions of sorghum and sprouted soybean in the formulation as evident in moisture content decrease with increase in sprouted soybean and increase with increase in sorghum (Table 1) in all the samples. Moisture content decreased due to sprouted soybean

may be attributed to soybean protein which binds and holds water (Igyor *et al.*, 2011). Variations in amylase and amylopectin proportions of sorghum may also contribute to the difference. Amylase is more soluble in water than amylopectin [30]. Moisture content similarities between samples 104 (75% sorghum: 25% sprouted soybean) and 106 (50% sorghum and 50% sprouted soybean) may mean that their blend variations did not have any significant ($p < 0.05$) different impart. Despite high MC of *akamu* wet paste still keep for some times on the shelf which can be attributed to fermentation that took place during 72h steeping thereby reduced microbial substrates.

5.1.3. Crude Protein Content

Protein is an energy substrate which is crucial for regulation and maintenance of infants' and young children's body [31]. Crude protein content obtained in this study ranged from 5.11% in sample 101 (100% sorghum) to 39.96% in sample 108 (25% sorghum: 75% soybean). And will meet the recommended protein content for complementary foods of 13g/d for infants aged from 0 to 2 years [32] by consuming 254g/d of paste from 100% sorghum while that from 25% sorghum: 75% soybean is an excellent protein source as it could meet the protein RDI per serving [33]. This is possible considering infant stomach capacity of 200ml [34]. Besides, both gruel samples are liable to prevent marasmus and kwashiorkor as well. Therefore, significant increase in protein content recorded in the paste could be traced to sprouted soybean [35] which contained higher proteins than sorghum (cereals). Increase in protein content of the paste with increase in soybean substitution was validated by the significant ($p < 0.05$) difference between samples 101 (100% sorghum) and 108 (25% sorghum: 75% soybean).

5.1.4. Crude Fat Content

Crude fat content of the samples increased with increase in soybean inclusion in the blends as shown in the significant higher fat content in sample 108 (25% sorghum: 75% soybean) than sample 101 (100% sorghum). This could be attributed to soybean which is an oil rich legume as can be visualized in the increasing fat content with increase in sprouted soybean inclusion in the table. Also significant ($p < 0.05$) fat content variation between all the samples substantiated significant ($p < 0.05$) difference in fat content contributions of various soybean proportions in the blends. Besides, all the pudding samples from both varieties with same formulations were significantly difference ($p < 0.05$) from each other. Fat contribution by the sprouted soybean harmonized with FAO/WHO recommendations of vegetable oil inclusion in infant and children foods [36] to increase the energy density and transport vehicle for fat soluble vitamins. Soybean contains considerable amount of polyunsaturated oil, omega-3 fatty acid and linolenic fatty acids which are good heart health nutrients for infants, children and adults [37]. Fat is a good source of energy, fat soluble vitamins and helps to increase food palatability as it absorbs and retains flavours [38].

5.1.5. Crude Fiber

Crude fiber contents of the paste increased from 1.41% in sample 101 (100% sorghum) to 4.82% in sample 108 (25% sorghum: 75% soybean) which implied that sprouted soybean was responsible for crude fiber increase as shown in the table. Also, DM increase with sprouted soybean inclusion earlier reported may have contributed too. Significant difference ($p < 0.05$) between all the samples may imply significant ($p < 0.05$) variations in fiber contributions of various blends.

Consumption of 1.04 to 3.55g/d of gruel samples will meet fiber RDI of 5g/d [39] for infants aged between 6 months to one year which made them a good fiber source [40]. Crude fiber is a carbohydrate subtype consisting of soluble and insoluble portions which increases roughage content of diet responsible, normalizing infant's bowel movement, prevents constipation, aids easier evacuation of stool and helps control blood pressure [39]. Dietary fiber helps in prevention of several diseases such as cancer, constipation, diabetes and intestinal diverticulitis [41, 42].

5.1.6. Ash Content

Ash content of the paste/samples which increased from 0.46% in sample 101 (100% sorghum) to 4.61% in sample 108 (25% sorghum: 75% soybean) implicated sprouted soybean inclusion levels in the formulation. Ash increase with sprouted soybean increase is evident in the table. Significant difference ($p < 0.05$) between all the samples may imply significant ($p < 0.05$) ash content increase by each soybean inclusion level. This may as well confirm higher ash content in sprouted soybean than sorghum. As ash is an index of mineral content of meal, sprouted soybean will go a long way in enhancing the mineral content of *akamu* successfully to meet the mineral need of the weaning infants.

5.1.7. Carbohydrate Content

Carbohydrate content of the paste which increased from 26.07% in sample 108 (25% sorghum: 75% soybean) to 77.44% in sample 101 (100% sorghum) implies that sorghum had more carbohydrate than soybean. Carbohydrate content of the paste decreased with increase in sprouted soybean inclusion in the blends as recognized in the table. Significant ($p < 0.05$) difference between all the paste samples again may signify significant variations in carbohydrate contributions of various blends. Soybean contains low carbohydrate (sucrose, starch and raffinose). Steeping and sprouting reduce starch and raffinose while sucrose is hydrolyzed into simple sugar [43]. Carbohydrates provide energy needed to fuel children's metabolism, support growth, keep their brain and nervous systems working and maintain overall health [44]. All the sprouted soybean blended pastes were good sources of carbohydrate despite their lower carbohydrate contents compared to 100% sorghum. Consumption of 2.30 to 3.64g/d of paste from sprouted soybean blended paste and 0.78 to 1.23g/d of paste from 100% sorghum will meet the recommended daily carbohydrate intake (RDI) of 60 to 95g/d for infants aged between 6 to 12

months [32].

5.1.8. Energy Values

Least energy value (335.26 Kcal) of sample 101 (100% sorghum) could be traced to least protein content due to absence of sprouted soybean. Sample 106 (50% sorghum: 50% sprouted soybean) had the highest energy value (364.9) 9K) than the rest samples probably due to higher contributions of all its energy substrates which ranked second except in carbohydrate. Protein, fat and crude fiber were enhanced by sprouted soybean (Table 1). Significant ($p<0.05$) energy difference between all the samples still point to the influence of sprouted soybean in the formulations.

5.2. Functional Properties

Results of functional properties of flour blends of sorghum and sprouted soybeans were shown in Table 2.

Table 3. Functional Properties of Sorghum and Sprouted Soybean flour Blends.

Products	Bulk density (g/ml)	Viscosity (μ Pas)	Swelling power	G-temperature ($^{\circ}$ C)	G-time (seconds)
101	0.33 ^b \pm 0.00	117.02 ^a \pm 0.01	22.83 ^a \pm 0.03	66.00 ^b \pm 0.00	0.35 ^c \pm 0.00
104	0.50 ^a \pm 0.00	109.04 ^b \pm 0.02	24.72 ^a \pm 0.01	65.50 ^{bc} \pm 0.50	0.36 ^c \pm 0.01
106	0.61 ^c \pm 0.00	102.02 ^b \pm 0.02	27.04 ^b \pm 0.01	49.00 ^c \pm 0.00	0.50 ^a \pm 0.01
108	0.66 ^c \pm 0.00	84.09 ^a \pm 0.01	30.04 ^a \pm 0.01	45.00 ^c \pm 0.00	0.57 ^a \pm 0.01

Values are mean duplicate determinations \pm standard deviation. Values with same superscript within same column are not significantly different ($p>0.05$). Values with different superscript within same column are significantly different. 101=100 % sorghum, 104=75% sorghum: 25% sprouted soybean, 106=50% sorghum: 50% sprouted soybean and 108=25% sorghum: 75% sprouted soybean.

5.2.2. Viscosity

Viscosity of the flour blends was highest (117) in sample 101 (100% sorghum) and least (84) in 108 (25% sorghum: 75% sprouted soybean) with significant ($P<0.05$) different between all the samples. The difference between the two viscosity values could be attributed to sprouted soybean inclusion as shown in Table 3 where viscosity decreased with increase inclusion of sprouted soybean flour. Sprouting thins down viscosity as most of the carbohydrates were hydrolyzed during sprouting unlike 100% sorghum flour. Significant ($p<0.05$) variations between samples may mean significant ($p<0.05$) variations in viscosity contributions by the blends. Viscosity is an indication of viscous gel after cooling and resistance to shear forces during shearing [46].

5.2.3. Swelling Power

Swelling power of the dried paste samples was highest (30.04) in sample 108 (25% sorghum: 75% sprouted soybean) and least (22.83) in sample 101 (100% sorghum) with significant ($p<0.05$) difference between all the samples. The difference could be attributed in part to the type of sorghum starch granules as well as inclusion of sprouted soybean which increased with increase in swelling power. Swelling power had been reported as indication of water holding capacity of starch and is generally used to differentiate between various types of starches [47, 48]. Water binding capacity in commercial starches stabilizes the quality and texture of some food products against syneresis during processing, storage and freezing. Besides, swelling

5.2.1. Bulk Density

Bulk density ranged from 0.33 in sample 101 (100% sorghum) to 0.66 in sample 108 (25% sorghum: 75% sprouted soybean) with significant ($p<0.05$) difference between all the samples which may be attributed to significant ($p<0.05$) variations in bulk density contributions of the various blends. Highest bulk density value of sample 108 (25% sorghum: 75% sprouted soybean) than the least value of sample 101 (100% sorghum) may mean that sprouted soybean paste contributed more bulk density than sorghum. This is substantiated by the increasing bulk density values of the samples with increasing substitution of sprouted soybean paste in all the samples. Bulk density depends on the intensity of attractive inter-particle forces, particle sizes as well as the number of contact points [45]. Hydrolysis of soybean due to sprouting which results in smaller particle sizes with larger surface areas may have favoured increase in bulk density of sprouted soybean.

power is related to protein and starch contents [49]. Therefore, soybean paste inclusion may likely contribute to the stability of the *akamu* paste. Protein hydrolyses during sprouting of soybean may have enhanced the swelling power of sample 108 (25% sorghum: 75% sprouted soybean) as high product protein content will embed the starch granules within its stiff matrix and gradually reduces their access to water thereby restrict the swelling power [50]. Higher swelling power of sprouted soybean suggested higher amylopectin content which is primarily responsible for granule swelling. Low amylase content is associated with high swelling power due to low reinforcement in the internal network by amylase molecules [51]. Higher swelling power of sprouted soybean used implies higher digestibility of the gruel made from their blends as amylase is more resistant to digestion due to its highly packed helical structure [30].

5.2.4. Gelatinization Temperature

Gelatinization temperature of the samples increased from 49 $^{\circ}$ C in sample 106 (50% sorghum and 50% sprouted soybean) to 66% in sample 101 (100% sorghum). Significant ($p<0.05$) difference between all the samples implies that *akamu* paste formulated from 50% sorghum and 50% sprouted soybean will form gel at a significantly ($p<0.05$) lower temperature than the rest sample blends. This may be attributed to variety, particle size of starch granules, and the proportion of amylose and amylopectin content of sorghum-sprouted soybean blends. Besides, Lower gelatinization temperature of sample 106 (50% sorghum and 50% sprouted

soybean) could be due to sprouted soybean paste inclusion. Sprouting may have predigested the soybean paste [37] thereby lowered their gelatinization temperature as could be seen in the decreasing trend of gelatinization temperature with increasing sprouted soybean paste substitution in Table 3. This may mean that sprouted soybean paste used in the formulation is compatible and complimentary to the *akamu*.

5.2.5. Gelatinization Time

Samples gelatinization time increased from 0.35sec in sample 101 (100% sorghum) to 0.57sec in sample 108 (25% sorghum: 75% sprouted soybean) with similarity between only samples 101 (100% sorghum) and 104 (75% sorghum: 25% sprouted soybean). The difference may be attributed to earlier report of higher amylopectin composition of sprouted soybean than sorghum starch which is larger in size and requires more time to gelatinize unlike in amylase. Therefore, inclusion of sprouted soybean increased the gelatinization time as evident in Table 3 where the gelatinization time increased with increasing substitution of sprouted soybean in the formulation. Similarity in gelatinization time between samples 101 (100% sorghum) and 104 (75% sorghum: 25% sprouted soybean) could be due to minimal (25%) substitution of sorghum with sprouted soybean which made no significant ($p < 0.05$) difference between them. Conversely,

significant ($p < 0.05$) difference between samples 106 (50% sorghum and 50% sprouted soybean) and 108 (25% sorghum: 75% sprouted soybean) could be due to higher substitution values (50% and 75% respectively) of sorghum with sprouted soybean.

5.3. Sensory Attributes

Sensory attributes of gruel prepared from sorghum-sprouted soybean formulated paste samples are presented in Table 4.

5.3.1. Appearance

Appearance scores of the gruel samples which ranged from 5.84 in sample 104 (75% sorghum: 25% sprouted soybean) to 7.30 in sample 101 (100% sorghum) may be attributed to inclusion of sprouted soybean which decreased appearance with increasing inclusion. Appearance score levels showed significant ($p < 0.05$) difference between samples 101 (100% sorghum) and the rest samples which similarities probably could be due to insignificant ($0 > 0.05$) effects on appearance by sprouted soybean inclusion levels. Appearance is an important sensory feature of any food product which decides acceptability as consumers eat with their eyes and use what they observed to predict quality [31].

Table 4. Sensory Attribute of Gruel Prepared from Sorghum and Sprouted Soybean Blends.

Product	Appearance	Taste	Thickness	Smoothness	G. acceptability
101	7.30 ^a ±1.34	6.20 ^{bc} ±1.10	8.25 ^a ±0.72	6.65 ^a ±0.67	7.10 ^a ±0.85
104	5.85 ^b ±1.87	4.60 ^{dc} ±1.87	5.25 ^{cd} ±2.02	6.00 ^a ±2.22	5.25 ^a ±2.2.9
106	6.00 ^{ab} ±2.07	2.90 ^f ±1.71	4.80 ^d ±2.04	6.25 ^a ±1.77	4.30 ^b ±2.00
108	6.00 ^{ab} ±1.95	4.45 ^e ±2.01	4.75 ^d ±2.40	6.75 ^a ±1.80	4.42 ^b ±2.04

Values are the means of the triplicate determinations±standard deviation, Values in same column with same superscripts are not significantly different ($p > 0.05$) and means with different superscripts are significantly ($p < 0.05$) different. Sample codes; 101=100% sorghum, 104=75% sorghum and 25% sprouted soybean, 106=50% sorghum: 50% sprouted soybean and 108=25% sorghum: 75% sprouted soybean.

5.3.2. Taste

Higher taste score (6.20) of gruel sample 101 (100% sorghum) than 2.90 from sample 106 (50% sorghum: 50% sprouted soybean) could be due to familiarity of the panelists with gruel from sorghum starch. Significant ($p < 0.05$) different between all the samples may prefigure significant ($p < 0.05$) variations in taste contributions of various levels of formulations as observed in the decreasing taste scores of the gruel samples with sprouted soybean inclusion levels compared to that of 100% sorghum gruel. Therefore, sprouted soybean inclusion should be with caution (less than 25%) to avoid total loss of acceptability. Probably that is why sample 106 (50% sorghum: 50% sprouted soybean) had the least taste score. Higher (6.20) taste score of 100% sorghum gruel may mean that taste attributes which include sweet, sour, bitter, salty, umami and other basic taste [52] were not influenced unlike in those formulated with sprouted soybean.

5.3.3. Thickness

Higher thickness value (8.25) of sample 101 (100% sorghum) than the least value (4.75) of sample 108 (25% sorghum: 75% sprouted soybean) is traceable to sprouted

soybean inclusion levels. Sprouting thins down (Iwe, 2010) the thickness of the gruel and hydrolyzes the native starch to simple sugar thereby reducing their thickening power. This is evident in the decreasing trend of the gruel thickness with increasing sprouted soybean inclusion in Table 4. Similarity between samples 106 (50% sorghum: 50% sprouted soybean) and 108 (25% sorghum: 75% sprouted soybean) could stem from the difference (25%) in their sprouted soybean inclusion which never had any significant ($p < 0.05$) thickness effect.

5.3.4. Smoothness

There were no significant ($p > 0.05$) difference between the smoothness of all the gruel samples. The similarities could be attributed to varieties of sorghum and soybean (granule sizes), steeping and sprouting which probably never had significant ($p < 0.05$) variations in the gruel smoothness. While steeping softens the starch, sprouting hydrolyzed the starch to simple sugars, both of which may have enhanced gruel smoothness equally. The smoothness scores increased from 6.00 in sample 104 (75% sorghum: 25% sprouted soybean) to 6.75 in sample 108 (25% sorghum: 75% sprouted soybean). The variation may be likened to the amount of sprouted soybean in the both blends. Sample 108 (25% sorghum: 75% sprouted

soybean) had more of sprouted soybean (50%) inclusion than 104 (75% sorghum: 25% sprouted soybean) which had more sorghum (50%). The difference may have been responsible for their significant ($p < 0.05$) difference. Thickness is an index is grittiness which in turn decides ease of swallowing. All the gruel samples are generally smooth which signified absence of swallowing problems.

5.3.5. General Acceptability

Sample 101 (100% sorghum) scored highest (7.10) by the panelists probably because they were used to gruel prepared from 100% sorghum. The least accepted was sample 106 (50% sorghum: 50% sprouted soybean) may be due to level of sprouted soybean inclusion (above 25%). Similarities of samples 104 (75% sorghum and 25% sprouted soybean) to 101 (100% sorghum), and 108 (25% sorghum: 75% sprouted soybean) to 106 (50% sorghum: 50% sprouted soybean) implied that general acceptability of the gruel decreased with increase in sprouted soybean inclusion most especially beyond 25%. Also, it may mean that variations in their formulations had no significant ($p < 0.05$) difference contributions in general acceptability. General acceptability depends on combination of all other sensory parameters and any product with maximum acceptability levels in most of the attributes will have maximum overall acceptability [53]. Maximum acceptability score (7.10) was justified with maximum scores in all the attributes evaluated. Following suit is gruel from sample 104 with maximum scores in taste and thickness, second in appearance and third in smoothness.

6. Conclusions

Supplementation of sorghum with sprouted soybean in *akamu* preparation is compatible as different proportions of the formulations contributed significant increase in protein, fat, ash, crude fiber and energy, and lowered carbohydrate contents. Some of the functional properties of the supplemented gruel samples like bulk density, swelling power and gelatinization time were also increased while viscosity and gelatinization temperature were decreased. Sensory evaluation revealed only increase in smoothness while appearance, taste, thickness and general acceptability were reduced most especially at inclusion level of sprouted soybean above 25%. Therefore, supplementation should be with caution, not more than 25%, to avoid loss of acceptability and thickness. Soy-*akamu* is therefore a good candidate for complementary feeding, breakfast meal for elderly people and those recovering from sickness to combat hidden hunger and protein energy malnutrition (PEM) problems among children of developing countries.

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