

Therapy Against COVID-19: Medicinal Plant Extracts Can Be a Solution

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Abstract: Coronavirus disease 2019 (COVID-19) was declared by the World Health Organization (WHO) as a pandemic on March 11, 2020. Despite the development of anti-COVID-19 vaccine, the disease continues to kill thousands of people, mainly due to a lack of sufficient doses for all populations and to the prioritization of populations to be vaccinated. According to epidemiological data from April 06, 2022, the COVID-19 pandemic has already killed more than 6,184,299 people around the world with nearly 494,286,073 confirmed cases. To date, no antiviral drug has been officially approved to fight this pandemic. Medical professionals and researchers all over the globe are in pursuit for the discovery therapies against this disease. One of the strategies is the use of medicinal plants. Indeed, plant extracts can inhibit viral growth and their effectiveness has been demonstrated on many viruses encountered in human pathology including SARS-CoV-1, poliovirus, varicella-zoster virus, HIV, human papillomavirus, HSV-1, HSV-2, influenza virus, cytomegalovirus and many others. Besides, it has been shown that plant extracts can regulate immune system and make the body able to prevent the establishment of viral infection or to fight against its deleterious effects. The exploration of plant extracts with anti-viral and immunostimulatory properties could be exploitable in the development of drug against COVID-19. This review promotes medicinal plant extracts as potential anti-COVID-19 drugs.

Keywords: COVID-19, Drug Research, Medicinal Plants

1. Introduction

Coronavirus disease 2019 (COVID-19) was declared a Public Health Emergency of International Concern by the World Health Organization (WHO) on 30 January 2020. The disease gradually spread across the world and was subsequently declared a pandemic on March 11, 2020. According to epidemiological data from January 25, 2021, the COVID-19 pandemic has already killed more than 6,184,299 people around the world with nearly 494,286,073 confirmed cases. It is an infectious disease with severe acute respiratory syndrome caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), a new type of coronavirus first discovered in the city of Wuhan, China, at the end of December 2019. The SARS-CoV-2 infects the

lower respiratory system induce a severe pneumonia. It can develop other disorders in the digestive system, heart, liver, kidneys, and nervous system, causing multi-organ failure. COVID-19-infected pneumonia is characterized by flu-like symptoms including fever, cough, and severe acute respiratory distress syndrome [1-4].

Since the outbreak of COVID-19, there has been no specific antiviral medication for treatment of the infection where supportive care and prevention of complications is the current management strategy. Despite the approval of the anti-COVID-19 vaccine by WHO, the disease continues to kill thousands of people, mainly due to a lack of sufficient doses for all populations and to the prioritization of

populations to be vaccinated. No antiviral drug is available to fight COVID-19 although there are many efforts around the world to develop one as quickly as possible. Enormous efforts have been made to protect, alleviate and cure the disease, though no specific treatment has been approved. Many countries infected by this virus, have done various proceedings such as distraction of traveling, isolation of patients and continuous medical care to reduce and prevent coronavirus spreading. The lack of treatment options for COVID-19 has led to many attempts to find alternative options to prevent the transmission of the disease or to alleviate the progression of the infection, including focusing more on preventive measures and the use of natural products and herbal extracts to increase immunity and decrease the probability of getting infected. Historically, the use of natural products was known to be the only treatment method against many infections and diseases. Medicinal plant species can provide a solution as a source of natural antiviral compounds by the accumulation of secondary metabolites and lectins as well as acting as a platform to express the viral immunogenic proteins [6-10].

The elderly patients or people who have existing chronic medical conditions such as heart failure, cancer, type 2 diabetes, severe obesity, chronic kidney disease, sickle cell disease, weakened immune system from solid organ transplants have a higher risk of serious illness from COVID-19. This can be due to the lower immune system. In order to combat SARS-CoV-2, many drug and vaccine development studies at experimental and clinical levels are currently conducted worldwide. Some traditional herbal medicines prevented SARS-CoV-2 infection of healthy persons and improved the health state of patients with mild or severe symptoms. Herbal drugs can effectively relieve symptoms, such as fever, cough, and fatigue, and reduce the probability of patients developing severe conditions. The contribution of traditional medicine in the management of Covid-19 may complement healthcare prevention and medical care services [14]. In China for example, the National Health Commission of China had recommended to use Chinese herbal medicine like QingfeiPaidu Decoction (QPD) to treat COVID-19 patients. Medicinal plants such as *Artemisia annua* are being considered by WHO as possible treatments for COVID-19 and should be tested for efficacy and adverse side effects. Furthermore, medicinal herbs have become an essential purpose to determine, isolate and purify the natural compounds for treatment of viral diseases. Most recent in silico studies revealed that natural compounds like myricitrin, methyl rosmarinate, calceolarioside B, myricetin3-O- β -D-glucopyranoside, licoleafol, and amaranthin could be potential leads to develop novel anti-SARS-CoV-2 drugs. WHO recognises that traditional, complementary and alternative medicine has many benefits. Medicinal plants have great prospect in the ultimate search for the cure against the dreaded coronavirus [2, 11-20].

The traditional medicinal system based on herbal therapies has always played a pivotal role in the health systems [20]. This review presents herbal remedies as a potential drug solution against COVID-19.

2. Plants with Antiviral and Immunomodulatory Properties

To date, medical professionals and researchers all over the globe are in pursuit for the discovery of prophylactic or anaphylactic remedy against COVID-19. One of the strategies employed is the exploration of traditional medicine as a potential source of an effective anti-COVID-19 drug. Plant extracts have long been used in the treatment of viral infections and their efficacy has been the subject of numerous scientific publications. Indeed, plant extracts can via several mechanisms inhibit viral growth and their effectiveness has been demonstrated on many viruses encountered in human pathology including SARS-CoV-1, poliovirus, varicella-zoster virus, HIV, human papillomavirus, HSV-1, HSV-2, influenza virus, cytomegalovirus and many others. By their antiviral properties, these plant extracts could also act on SARS-CoV-2, the viral agent responsible for COVID-19. It has also been shown that plant extracts can act at several levels of the body and regulate various mechanisms including the expression of the immune system. As with any other microbial infection, the immune system plays an important role in controlling COVID-19 infection. The immunocompromised are the most at risk. By boosting the immune system, plant extracts with immunostimulating properties make the body able to prevent the establishment of infection or to fight against its deleterious effects. Plant extracts can enhance immune reactions via the stimulation of non-specific systems, such as granulocytes, macrophages, complement, certain T-lymphocytes and various effector substances. The selection of plant extracts proven to be safe and possessing anti-SARS-CoV-2 and immunostimulatory properties could be exploitable in the development of an effective anti-COVID-19 drug. Plant extracts provide a rich resource for novel antiviral drug development and can improve the immunity against viruses or other pathogen. In several countries around the world, drug formulations made from medicinal plants are used to prevent and fight COVID-19. This is the case with Qing FeiPai Du Tang (QFPDT), a Chinese medicine formula which has been reported to be efficacious on COVID-19 in China. The main herbs of QFPDT have been shown to have antiviral effects via different mechanisms. It can direct effect on virus replication and autophagy, and promotes the human defense system via T and B cell functions [21-23]. Table 1 below presents some plants with antiviral which can also act on immune system.

Table 1. Plants with antiviral and immunomodulatory properties.

Medicinal plants/family	Susceptible viral species	Effect on the immune system	References
<i>Acacia nilotica</i> (L.) Willd. ex Delile / Fabaceae	Influenza A virus, hepatitis C Virus	Modulates splenocyte proliferation and IL-10 secretion	[24-26]
<i>Aloe vera</i> (L.) Burm. F. / Aloeaceae	Coronavirus SARS-CoV-1, poliovirus, varicella-zoster virus, HIV, human papillomavirus, haemorrhagic viral rhobdavirus septicaemia, Herpes simplex virus type 1, Herpes simplex virus type 2, influenza virus, cytomegalovirus	Stimulates humoral immunity, activates macrophages and cytokine release, stimulates cellular immune response and prevents immune suppression, stimulates antibody production	[27-33]
<i>Andrographispaniculata</i> (Burm. f.) Wall. ex Nees / Acanthaceae	HIV, flaviviruses, pestiviruses, herpes simplex virus, Epstein-Barr virus	Induces lymphocyte proliferation, inhibits the production of TNF- α and IL-12 by macrophages, stimulates the production of antibodies, improves the tolerogenic properties of immature dendritic cells, stimulates the innate immune response, increases the production of IL-2, reduces the secretion of IFN- γ and IL-2 in murine T cells	[34-39]
<i>Annonamuricata</i> L. / Annonaceae	Dengue virus type 2	Induces transcriptional expression of TNF- α and IL-1 β via MAP kinase pathways, enhances the immune activity of the RAW 264.7 macrophage, increases white blood cells, T lymphocytes and natural killer cells, promotes the production of nitric oxide, IL-6 and TNF- α , modulates effector functions of human macrophages	[40-43]
<i>Artemisia annua</i> L. / Asteraceae	Herpes simplex virus type 1, bovine viral diarrhoea virus	Stimulates the production of Th1 cytokines (IFN- γ) and inhibits that of the Th2 pathway (IL-4 and IL-10), increases the levels of CD4+ and CD8+ T cells, stimulates lymphoproliferation, increases co-stimulating molecules CD80 and CD86 on APCs, stimulates the generation of NO and enhances the Th1 immune response	[44-47]
<i>Azadirachtaindica</i> A. Juss. / Meliaceae	Group B Coxsackie viruses, duck plague virus	Increases serum IL-2 level and induces proliferation of T lymphocytes in thymus, increases IgM and IgG level, improves phagocytic activity of immune cells, increases red blood cells, white blood cells, lymphocytes, monocytes and neutrophils, decreases IgE and IL-4 expression, has anticomplement activity, activates cell-mediated immune mechanisms, stimulates PMNs, activates macrophages, increases phagocytosis and reduction of NBT, promotes proliferative responses of T lymphocytes, stimulates the secretion of IFN- γ by T cells	[48-54]
<i>Boerhaviadiffusa</i> L. / Nyctaginaceae	Potato virus X, mung bean yellow mosaic virus	Stimulates cell-mediated immune response by upregulating IL-2 and downregulating pro-inflammatory cytokines such as IL-1 β , IL-6 and TNF- α , enhances NK cell activity, antibody-dependent cell cytotoxicity and antibody-dependent complement-mediated cytotoxicity, enhances leukocyte production, enhances phagocytic activity of macrophages	[55-57]
<i>Carissa edulis</i> (Forssk.) Vahl / Apocynaceae	Herpes simplex virus	Induces an increase in red blood cells, lymphocytes, monocytes, neutrophils, eosinophils and basophils	[58, 59]
<i>Echinacea purpurea</i> (L.) Moench / Asteraceae	Coronavirus, Herpes simplex virus type 1, respiratory syncytial virus, influenza A virus, influenza B virus	Improves NK cell activity and B cell response, increases T cell proliferation and cytokine production, increases IgG, interferon γ and cytokine expression, increases hemoglobin level, red blood cells, white blood cells, lymphocytes, IgM and stimulates phagocytosis, increases the functions of NK cells, p38 MAPK, NF-kB, JNK, differentiation of DC and phagocytic and bactericidal activity intracellular, stimulates production IL-1, IL-6, IL-12p70, TNF- α , NO, CD80, CD86 and MHCII, activates macrophages, CCR7, IL-10, PPAR- γ , IL-2, IFN- γ , reduces number and function of regulatory T cells	[60-70]
<i>Eclipta alba</i> Hassk / Asteraceae	Hepatitis C Virus, HIV	Increases the phagocytic index of PMNs, antibodies and white blood cells	[71, 72]
<i>Jatropha curcas</i> L. / Euphorbiaceae	HIV, Newcastle disease challenge virus	Stimulates both humoral and cell-mediated seroresponse, increases antibody, lymphocyte and macrophage cells	[73, 74]
<i>Kigelia Africana</i> (Lam.) Benth. / Bignoniaceae	Herpes Simplex Virus, HIV	Increases both humoral immunity and cell mediated immunity, increases antibody level	[75, 76]
<i>Emilia sonchifolia</i> (L.) DC / Asteraceae	White spot syndrome virus, yellow head virus, japanese encephalitis virus,	Increases white blood cell count, bone marrow cellularity, α -esterase activity and lymphoid organ weight, increases antibody titer, increases cell-mediated immune response by enhancing the activity of destruction of CTL, improves production of IL-2 and IFN- γ	[77-79]
<i>Mangiferaindica</i> L. / Anacardiaceae	Herpes simplex virus type 2, poliovirus type-1, influenza virus.	Increases humoral antibody (HA) titer, neutrophil adhesion, PMN phagocytic index, improves innate and adaptive immune response via increased blood cell count, spleen index and titer 'hemagglutination (HA)	[80-84]
<i>Moringaoleifera</i> Lam. / Moringaceae	HIV, herpes simplex virus type, HBV, EBV, FMDV, NDV	Stimulates both cellular and humoral immunity, increases WBC, neutrophils, neutrophil adhesion, antibody, lymphocytes, eosinophils, monocytes, weight of thymus and spleen, phagocytic index, increase the cell number of CD4+ and CD8+	[85-88]
<i>Ocimumbasilicum</i> L. / Lamiaceae	Adenoviruses, hepatitis B virus, coxsackievirus B1,	Stimulates lymphoproliferation, increases WBC	[89-91]

Medicinal plants/family	Susceptible viral species	Effect on the immune system	References
	herpes viruses, enterovirus 71, HIV-1		
<i>Phyllanthus niruri</i> L. / Phyllanthaceae	Hepatitis B virus, herpes simplex viruses, dengue virus, HIV	Increases polymorphonuclear neutrophils and leucocytes mobilization, leucocytes count, humoral antibody titre, induces proliferation of peripheral blood mononuclear cells, increases NO release, and improves macrophages phagocytic activity, increases WBC and lymphocytes	[92-95]
<i>Phyllanthus reticulatus</i> Poir. / Euphorbiaceae	Herpes simplex virus type 1, Herpes simplex virus type 2	Increases the phagocytic activity of mononuclear macrophage, neutrophil adhesion, and white blood cell, acts on NK cells by promoting NFκB signaling enhancement	[96-98]
<i>Psidium guajava</i> Linn. / Myrtaceae	Influenza viruses, dengue virus	stimulates both humoral and cell mediated immunity, increases the WBC, RBC, platelet, monocyte, neutrophil, eosinophil, lymphocyte, antibody and hemoglobin levels	[99-101]
<i>Solanum nigrum</i> L. / Solanaceae	Hepatitis C Virus	Increases the antibody response, IgG level, neutrophil activity, induces phagocytosis activity and stimulated the production of TNF-α and IL-6, activates macrophages, increases IFN-α, IL-2 and IFN-γ production	[102-105]
<i>Tinospora cordifolia</i> (Thunb.) Miers / Menispermaceae	HIV virus, hepatitis-A Virus	Increases the phagocytic activity of macrophages, the production of reactive oxygen species (ROS) in human neutrophils, improves the production of nitric oxide (NO) by stimulation of splenocytes and macrophages, stimulates the production of cytokines, mitogenicity, the activity of immune effector cells, regulates the activity of IL-6 cytokines, activates cytotoxic T lymphocytes and B lymphocyte differentiation, induces IL-1 secretion, activates lymphocytes and synthesis of pro and anti-inflammatory cytokines	[106]
<i>Vernonia amygdalina</i> Del./Asteraceae	HIV, hepatitis B virus	Increase CD4+ T cell, IgM, IgG1 and IgA antibody responses, white blood cell, neutrophil and lymphocyte counts, decreases IL-6 level	[107-109]
<i>Zingiber officinalis</i> L. / Zingiberaceae	Chikungunya virus, human respiratory syncytial virus, hepatitis C virus	Stimulates phagocytic activity of white blood cells, humoral immune response, increases hematocrit, red blood cells and white blood cells	[110-113]

3. Conclusion

The advent of the COVID-19 pandemic has totally changed global habits both economically and socially. To date, no antiviral drug has been approved to fight this disease. Traditional medicine once held pride of place as the premier treatment for emerging diseases. Several plant species are known there for their therapeutic and anti-infectious properties in particular. Since the advent of COVID-19 many potions made from herbal remedies have been proposed as drugs to fight this infection. However, no scientific studies demonstrating the efficacy and safety of these drugs for approval have been performed. Clinical studies on these drug potions and an exploration of other therapeutic possibilities offered by traditional medicine would help to overcome this pandemic.

References

- [1] Worldometer. COVID-19 Coronavirus Pandemic. April 06, 2022.
- [2] Laksmiani NP, Luh PF, Anak AG, Putu AA, Anak AI, and Ni Putu AK. Active Compounds Activity from the Medicinal Plants against SARS-CoV-2 using in Silico Assay. *Biomedical & Pharmacology Journal*. 2020; 13 (2): 873-881.
- [3] Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020; S0140-6736 (0120): 30183-5.
- [4] Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, Qiu Y, Wang J, Liu Y, Wei Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*. 2020.
- [5] Eman S, Ahmed AN, Reham SI. Potential role of medicinal plants and their constituents in the mitigation of SARS-CoV-2: identifying related therapeutic targets using network pharmacology and molecular docking analyses. *The Royal Society of Chemistry*. 2020; 10: 27961-27983.
- [6] Spinney L. "When will a coronavirus vaccine be ready?". *The Guardian*. 2020.
- [7] Yonesi M, Rezazadeh A. Plants as a Prospective Source of Natural Anti-viral Compounds and Oral Vaccines against COVID-19 Coronavirus. Preprints 2020. doi: 10.20944/preprints202004.0321.v1.
- [8] Ahn D, Shin H, Kim M, Lee S, Kim H, Myoung J, Kim B, Kim S. Current Status of Epidemiology, Diagnosis, Therapeutics, and Vaccines for Novel Coronavirus Disease 2019 (COVID-19). *J Microbiol Biotechnol*. 2020; 30 (3): 313-324.
- [9] Ali AA, Mustafa FH. Perspective Study of Exploring Some Medicinal Plants to Manage the Pandemic COVID-19. *European Journal of Medical and Health Sciences*. 2020; 2 (4): 1-5.
- [10] Hamad SA, Mohamed AA, Fahad MA, Hamad NA, Wejdan IA, Abdulrahman IA, Abdullah SA, Rawan AA, Njood AA. Knowledge about COVID-19 and patients' beliefs about and use of herbal products during the COVID-19 pandemic: a cross-sectional study in Saudi Arabia. 2020.
- [11] Mayo C. Coronavirus disease 2019 (COVID-19). 1998-2020 Mayo Foundation for Medical Education and Research (MFMER). 2020.

- [12] Ilkay EO, Sezer SD. Natural Products as Potential Leads Against Coronaviruses: Could They be Encouraging Structural Models Against SARS-CoV-2? *Natural Products and Bioprospecting*. 2020; 10: 171-186.
- [13] Hong-Zhi DU, Hou XY, Miao YH, Huang BS, Liu DH. Traditional Chinese Medicine: an effective treatment for 2019 novel coronavirus pneumonia (NCP). *Chin. J. Nat. Med.* 2020; 18 (3): 226-230.
- [14] Sanogo R. Traditional medicine resources in the fight against Covid-19. Friedrich-Ebert-Stiftung. 2020.
- [15] National Health Commission. Press conference of the joint prevention and control mechanism of the State Council on April 14. 2020. <http://www.nhc.gov.cn/xcs/fkdt/202004/05f7>
- [16] Jin YH, Cai L, Cheng ZS, Cheng H, Deng T, et al. A rapid advice guideline for the diagnosis and treatment of 2019 novel coronavirus (2019-nCoV) infected pneumonia (standard version). *Military Medical Research*. 2020; 7: 4.
- [17] World Health Organization (WHO). WHO supports scientifically-proven traditional medicine. World Health Organization, Regional Office for Africa. 2020.
- [18] Dhama K, Karthik K, Khandia R, Munjal A, Tiwari R, Rana R. Medicinal and Therapeutic Potential of Herbs and Plant Metabolites/Extracts Countering Viral Pathogen-Current Knowledge and Future Prospects. *Curr Drug Metab.* 2018; 19 (3): 236-263.
- [19] Erdogan OI, Sezer FS. Natural Products as Potential Leads Against Coronaviruses: Could They be Encouraging Structural Models Against SARS-CoV-2? *Natural Products and Bioprospecting*. 2020; 10: 171-186.
- [20] Onah EO. Medicinal plants: Prospective drug candidates against the dreaded Coronavirus. *Iberoamerican Journal of Medicine*. 2020; 04: 314-321.
- [21] Al-Khafaji K, Al-Duhaidahawi LD, Taskin TT. Using Integrated Computational Approaches to Identify Safe and Rapid Treatment for SARS-CoV-2. *Journal of Biomolecular Structure and Dynamics*. 2020; 1-11.
- [22] Sanghai VD, Kulkarni SR, Sanghai NN. Screening Of Antiviral Compounds From Plants. *Journal of Pharmacy Research*. 2014; 8 (8): 1050-1058.
- [23] Li DZ, Wai CL, Wei Y, Kam WC, Stephen CW, Jiangxia M, Ken KL, Zhaoxiang B, Vivian TW. Potential Targets for Treatment of Coronavirus Disease 2019 (COVID-19): A Review of Qing-Fei-Pai-Du-Tang and Its Major Herbs. *The American Journal of Chinese Medicine*. 2020; 48 (5): 1051-1071.
- [24] Mona TI, Malik SM, Sarawut K, Natthida T, Duncan RS, Abdurahman NH, Azhari NH, Alamin IE. Antiviral activity and possible mechanisms of action of *Acacia nilotica* against Influenza A virus. *J Clin Cell Immunol*. 2016.
- [25] Sidra R, Usman AA, Sana R, Tariq J, Sheikh R. Antiviral activity of *Acacia nilotica* against Hepatitis C Virus in liver infected cells. *Virology Journal*. 2011; 8 (1): 220.
- [26] Arvind KS, Amit K, Sharad KY, Anu R. Studies on Antimicrobial and Immunomodulatory Effects of Hot Aqueous Extract of *Acacia nilotica* L. Leaves against Common Veterinary Pathogens. *Veterinary Medicine International*. 2014; 9.
- [27] Mpiana PT, Koto-Te-Nyiwa N, Damien ST, Jason TK, Benjamin ZG, Domaine TM, Inkoto CL, Lengbiye EM, Mbadiko CM, Matondo A, Bongo GN, Tshilanda DD. *Aloe vera* (L.) Burm. F. as a Potential Anti-COVID-19 Plant: A Mini-review of Its Antiviral Activity. *European Journal of Medicinal Plants*. 2020; 31 (8): 86-93.
- [28] Sumita H, Ashish KM, Pramod KM. Augmented humoral immune response and decreased cell-mediated immunity by *Aloe vera* in rats. *Inflammopharmacology*. 2012; 20 (6).
- [29] Zohreh F, Tooba G, Roya Y. Immunomodulatory effects of *Aloe vera* and its fractions on response of macrophages against *Candida albicans*. *Immunopharmacology and Immunotoxicology*. 2011; 33 (4): 676-81.
- [30] Pugh N, Ross SA, ElSohly MA, Pasco DS. Characterization of aloeride, a new high-molecular-weight polysaccharide from *Aloe vera*. with potent immunostimulative activity. *J Agric Food Chem*. 2001; 49: 1030-1034.
- [31] Oronz-Barocio A, Zaitseva G, Chavez-Anaya A, Arceta-Gonzalez VI, Puebla-Perez AM, Alfaro-Bustamante F, Zimina IV, Arion VY. Modulation of immune response of BALB/Mice bearing lymphoma L5178Y treated with bitter yellow juice of *Aloe vera*. (L) *in vivo*. *Russ J Immunol*. 1999; 4: 43-50.
- [32] Yagi A, Takeo S. Anti-inflammatory constituents, aloesin and aloemannan in *Aloe* species and effects of tanshinon VI in *Salvia miltiorrhiza*. on heart. *YakugakuZasshi*. 2003; 123: 517-532.
- [33] Peng SY, Norman J, Curtin G, Corrier D, McDaniel HR, Busbee D. Decreased mortality in Norman murine sarcoma in mice treated with the immunomodulator, Acemannan. *MolBiother*. 1991; 3: 79-87.
- [34] Churiyah OP, Elrade R, Tarwadi. Antiviral and Immunostimulant Activities of *Andrographispaniculata*. *HAYATI Journal of Biosciences*. 2015; 22 (2): 67-72.
- [35] Thanasekaran J, Cheng-Ying H, Jie-Jen L, Joen-Rong S. Experimental and Clinical Pharmacology of *Andrographispaniculata* and Its Major Bioactive Phytoconstituent And rographolide. *Evidence-Based Complementary and Alternative Medicine*. 2013; 16.
- [36] Seubsasana S, Pientong TE, Thongchai S, Aromdee C. A potential andrographolide analogue against the replication of herpes simplex virus type 1 in vero cells. *Medicinal Chemistry*. 2011; 7 (3): 237-244.
- [37] King SL. Andrographolide derivatives to treat viral infections. 2006.
- [38] Qin LH, Kong L, Shi GJ, Wang ZT, Ge BX. Andrographolide inhibits the production of TNF- α and interleukin-12 in lipopolysaccharide-stimulated macrophages: role of mitogen-activated protein kinases. *Biological & Pharmaceutical Bulletin*. 2006; 29 (2): 220-224.
- [39] Iruetagoiena MI, Tobar JA, González PA et al. Andrographolide interferes with T cell activation and reduces experimental autoimmune encephalomyelitis in the mouse. *The Journal of Pharmacology and Experimental Therapeutics*. 2005; 312 (1): 366-372.
- [40] Wahab NZ, Ibrahim N, Kamarudin MK, Lananan F, Juahir H, Ghazali A, Ireana YA. Cytotoxicity and antiviral activity of *Annonamuricata* aqueous leaves extract against dengue virus type 2. *Special Issue*. 2018; 10 (1S).

- [41] Goon-Tae K, Nguyen Khoi ST, Eun-Hye C, YooJeong S, Jae-Hwi S, Soon-Mi S, Tae-sik P. Immunomodulatory Efficacy of Standardized *Annonamuricata* (Graviola) Leaf Extract via Activation of Mitogen-Activated Protein Kinase Pathways in RAW 264.7 Macrophages. Evidence-based Complementary and Alternative Medicine. 2016; 2: 1-10.
- [42] Syed NS, Romli MF, Hamid M, Alitheen NB, Abd R. Anticancer effect of *AnnonaMuricata* Linn Leaves Crude Extract (AMCE) on breast cancer cell line. BMC Complement Altern Med. 2016; 16: 311.
- [43] Adan J, Arana-Argaez V, Torres-Romero J, Canul-Canche J, Ramirez-Camacho M. Effect of methanolic extract of *Annonamuricata* leaves on macrophage effector functions. J Immunol. 2016; 196.
- [44] Mehrangiz KK, Seyed AE, Masoud SG, Sani EA, Sahebkar A. Antiviral activities of aerial subsets of Artemisia species against Herpes Simplex virus type 1 (HSV1) *in vitro*. Asian Biomedicine. 2011; 5 (1): 63-684.
- [45] Romero MR, Serrano MA, Vallejo M, Efferth T, Alvarez M, Marin JJ. Antiviral Effect of Artemisinin From *Artemisia Annua* Against a Model Member of the Flaviviridae Family, the Bovine Viral Diarrhoea Virus (BVDV). Planta Med. 2006; 72 (13): 1169-74.
- [46] Mohammad I, Garima C, Abdullah F, Bilikere SD, Dinkar S, Farhat A. Th1-Biased Immunomodulation and Therapeutic Potential of *Artemisia annua* in Murine Visceral Leishmaniasis. PLoSNegl Trop Dis. 2015; 9 (1): e3321.
- [47] Chenchen S, Haipeng L, Yifu Y, Lifei H. Anti-Inflammatory and Immunoregulatory Functions of Artemisinin and Its Derivatives. Anti-inflammatory Natural Products. 2015; 7.
- [48] BadamL, Swati PJ, Sabah B. *In vitro* antiviral activity of neem (*Azadirachtaindica*. A. Juss) leaf extract against group B Cocksackieviruses. The Journal of communicable diseases. 1999; 31 (2): 79-90.
- [49] Xu J, Song X, Yin ZQ, Cheng AC, Jia RY, Deng YX, Ye KC, Shi CF, Lv C, Zhang W. Antiviral activity and mode of action of extracts from neem seed kernel against duck plague virus *in vitro*. Immunology, Health, and Disease. 2012; 91 (11): 2802-2807.
- [50] Venugopalan SK, Visweswaran N, Aiyalu R, Narenk N, Dorai SP, Srinivasan N, Subramaniam R. Isolation and characterization of glucosamine from *Azadirachtaindica* leaves: An evaluation of immunostimulant activity in mice. Asian Pacific Journal of Tropical Biomedicine. 2012; 2 (3): S1561-S1567.
- [51] Ray A, Banerjee BD, Sen P. Modulation of humoral and cell-mediated immune responses by *Azadirachtaindica* (Neem) in mice. Indian J Exp Biol. 1996; 34 (7): 698-701.
- [52] Truong QN, Bui TB, Le TB, Quetin-Leclercq J, Scippo ML, Thanh P, Kestemont P. Plant extract-based diets differently modulate immune responses and resistance to bacterial infection in striped catfish (*Pangasianodonhypophthalmus*). Fish & Shellfish Immunology. 2019; 92: 913-924.
- [53] Ratna D, Susilawati, Pujiana EL, Roedy B, Erawati W, Ristya W, Sunlip W. Aqueous Extract of Neem Leaves (*Azadirachtaindica*) Decrease Expression of Immunoglobulin E (IgE) and Interleukin 4 (IL-4) in Gingiva Tissue of BALB/c Mice Injected by Ovalbumine. VMIC. 2017.
- [54] Njiro SM, Kafi-Tsekpo MW. Effect of an aqueous extract of *Azadirachtaindica* on the immune response in mice. Onderstepoort J. Vet. Res. 1999; 66: 59-62.
- [55] Ganesh KA, Sankarganesh P, Baby J. Antiviral Plant Extracts. IntechOpen. 2019; 1: 1-10.
- [56] Manu KA, Girija K. Effect of Punarnavine, an Alkaloid from *Boerhaaviadiffusa*, on Cell-Mediated Immune Responses and TIMP-1 in B16F-10 Metastatic Melanoma-Bearing Mice. Immunopharmacology and Immunotoxicology. 2007; 29 (3-4): 569-86.
- [57] Sumanth M, Mustafa SS. Antistress, adoptogenic and immunopotentiating activity roots of *Boerhaaviadiffusa* in mice. International Journal of Pharmacology. 2007; 3 (5): 416-420.
- [58] Mukhtar M, Mohammad A, Mahmood A, Pomerantz RJ, Wigdahl B, Parveene Z. Antiviral potentials of medicinal plants. Virus Res. 2008; 131 (2): 111-120.
- [59] Jorum OH, Ngugi MP, Machocho AK. Haematological Effects of Dichloromethane-Methanolic Leaf Extracts of *Carissa edulis* (Forssk.) Vahl in Normal Rat Models. Journal of Hematology & Thromboembolic Diseases. 2016; 4 (1): 1-8.
- [60] Hudson JB. Applications of the Phytomedicine *Echinacea purpurea* (Purple Coneflower) in Infectious Diseases. Natural Products for Medicine. 2011; 1-16.
- [61] Zili Z, Yi L, Lankun W, Senchina DS, Wurtele ES, Murphy PA, Kohut ML, Cunnick JE. Enhancement of Innate and Adaptive Immune Functions by Multiple *Echinacea* Species. J Med Food. 2007; 10 (3): 423-434.
- [62] Torkan S, Khamesipour F, Katsande S. Evaluating the effect of oral administration of *Echinacea* hydroethanolic extract on the immune system in dog. Autonomic and Autacoid Pharmacology. 2015; 35 (1-2): 9-13.
- [63] Cuneyt S, Gamze AK, Nuri T, Aysun Y, Utku YC, Ozge A, Juergen AR, Huseyin Y. Immunomodulatory effects of *Echinacea* and *Pelargonium* on the innate and adoptive immunity in calves. Food and Agricultural Immunology. 2018; 29 (1) 744-761.
- [64] See DM, Broumand N, Sahl L, Tilles JG. *In vitro* effects of *echinacea* and *ginseng* on natural killer and antibody-dependent cell cytotoxicity in healthy subjects and chronic fatigue syndrome or acquired immunodeficiency syndrome patients. Immunopharmacology. 1997; 35: 229-235.
- [65] Wang CY, Chiao MT, Yen PJ, Huang WC, Hou CC, Chien SC, Yeh KC, Yang WC, Shyr LF, Yang NS. Modulatory effects of *Echinacea purpurea* extracts on human dendritic cells: A cell and gene-based study. Genomics. 2006; 88: 801-808.
- [66] Fu A, Wang Y, Wu Y, Chen H, Zheng S, Li Y, Xu X, Li W. *Echinacea purpurea* extract polarizes M1 macrophages in murine bone marrow-derived macrophages through the activation of JNK: *Echinacea purpurea* extract polarizes M1 macrophages. J. Cell. Biochem. 2017; 118: 2664-2671.
- [67] Chicca A, Raduner S, Pellati F, Strompen T, Altmann KH, Schoop R, Gertsch J. Synergistic immunopharmacological effects of N-alkylamides in *Echinacea purpurea* herbal extracts. Int. Immunopharmacol. 2009; 9: 850-858.

- [68] Spelman K, Iiams-Hauser K, Cech NB, Taylor EW, Smirnoff N, Wenner CA. Role for PPAR in IL-2 inhibition in T cells by Echinacea-derived undeca-2E-ene-8,10-dienoic acid isobutylamide. *Int. Immunopharmacol.* 2009; 9: 1260-1264.
- [69] Fonseca FN, Papanicolaou G, Lin H, Lau CB, Kennelly EJ, Cassileth BR, Cunningham-Rundles, S. *Echinacea purpurea* (L.) moench modulates human T-cell cytokine response. *Int. Immunopharmacol.* 2014; 19: 94-102.
- [70] Catanzaro M, Corsini E, Rosini M, Racchi M, Lanni C. Immunomodulators Inspired by Nature: A Review on Curcumin and Echinacea. *Molecules.* 2018; 23 (2778): 1-17.
- [71] Hyung-Ran K, Sei-Kwan O, Woosung L, Hyeon KL, Byung-In M, Ju-Young S. Immune Enhancing Effects of *Echinacea purpurea* Root Extract by Reducing Regulatory T Cell Number and Function. *Natural Product Communications.* 2014; 9 (4): 511-514.
- [72] Razdan R, Imranulla A, Dev MJ. Preventive and curative effects of Vedic guard against antitubercular drugs induced hepatic damage in rats. *Pharmacognosy Magazine.* 2008; 4 (15): 182-188.
- [73] Jayathirtha MG, Shrihari HM. Preliminary immunomodulatory activities of methanolic extracts of *Ecliptaalba* & *Centellaasiatica*. *Phytomedicine.* 2004; 11 (4): 361-5.
- [74] Ritwik D, Soumen R, Deepak YP, Deshmukh RR et al. Potential Anti-HIV Activity of *Jatropha curcas* Linn. Leaf Extracts. *Journal of Antivirals and Antiretrovirals.* 2013; 5 (7): 160-165.
- [75] Howaida IA, Fatma M, Gaara AH, El-Safty MM. Phytoconstituents of *Jatropha curcas* L. Leaves and their Immunomodulatory Activity on Humoral and Cell-Mediated Immune Response in Chicks. *Zeitschrift fur Naturforschung C.* 2009; 64 (7-8): 495-501.
- [76] Rukanga GM, Kofi-Tsekpo MW, Kurokawa M, Kageyama S, Mungai GM, Muli JM, Tolo FM, Kibaya RM, Muthaura CN, Kanyara JN, Tukei PM, Shiraki K. Evaluation of the HIV-1 reverse transcriptase inhibitory properties of extracts from some medicinal plants in Kenya. *Afr J Health Sci.* 2002; 9 (1): 81-90.
- [77] Reena K, Nagarathna PK, Sriram R, Wesley J. Evaluation of Immunomodulatory activity of the flavanoid from *Kigelia Africana*. *Indian Journal of Pharmaceutical and Biological Research.* 2014; 2 (2): 41-48.
- [78] Gilcy GK, Girija K. Immune response modulatory effect of *Emilia sonchifolia* (L.) DC: an in vivo experimental study. *Journal of Basic and Clinical Physiology and Pharmacology.* 2015; 26 (6): 613-622.
- [79] Lamai M, Wilaiwan C, Wilawan M. *Emilia sonchifolia* extract activity against white spot syndrome virus and yellow head virus in shrimp cell cultures. *Dis Aquat Org.* 2015; 115: 157-164.
- [80] Yadava RN, Mamta R. Antiviral activity of a new flavone glycoside from *Emilia sonchifolia* DC. *Indian Journal Chemistry.* 2012; 15B: 635-638.
- [81] Zhu XM, Song JX, Huang ZZ, Wu YM, Yu MJ. Antiviral Activity of Mangiferin Against Herpes Simplex Virus Type 2 *in Vitro*. *Zhongguo Yao Li Xue Bao.* 1993; 14 (5): 452-4.
- [82] Rechenchoski DZ, Faccin-Galhardi LC, Pacheco CA, Ngila MP, Nozawa C, Carvalho LR. Antiviral potential of mangiferin against poliovirus. *Int J of Pharmc Res.* 2018; 8 (4).
- [83] Amin AS, Hajir SH, Marwa AA. Antiviral Activity of Mangifera Extract on Influenza Virus Cultivated in Different Cell Cultures. *J Pure Appl Microbiol.* 2019; 13 (1): 455-458.
- [84] Chetan S, Anand RK, Basheerahmed AM, Gajare R. Immunostimulant phytoconstituents from *Mangifera indica* L. bark oil. *The Journal of Phytopharmacology.* 2014; 3 (2): 139-148.
- [85] Endang K, Siti NA, Siti MA, Waqas A. Immunostimulant activity of standardised extracts of *Mangifera indica* leaf and *Curcuma domestica* rhizome in mice. *Tropical Journal of Pharmaceutical Research.* 2018; 17 (1): 77-84.
- [86] Biswas D, Nandya S, Mukherjee A, Pandey DK, Dey A. *Moringaoleifera* Lam. and derived phytochemicals as promising antiviral agents: A review. *South African Journal of Botany.* 2020; 129: 272-282.
- [87] Anamika G, Manish G, Rahul KS, Shampa A et al. Immunomodulatory effect of *Moringaoleifera* Lam. extract on cyclophosphamide induced toxicity in mice. *Indian journal of experimental biology.* 2010; 48 (11): 1157-60.
- [88] Nfambi J, Bbosa GS, Fred SL, Gakunga J, Kasolo JN. Immunomodulatory activity of methanolic leaf extract of *Moringaoleifera* in Wistar albino rats. *J Basic Clin Physiol Pharmacol.* 2015; 26 (6): 603-611.
- [89] Rachmawati I, Muhaimin R. *In vitro* Immunomodulatory Activity of Aqueous Extract of *Moringaoleifera* Lam. Leaf to the CD4+, CD8+ and B220+ Cells in Mus musculus. *J. Exp. Life Sci.* 2014; 4 (1): 15-20.
- [90] Lien-Chai C, Lean-Teik N, Pei-Win C, Win C, Chun-Ching L. Antiviral activities of extracts and selected pure constituents of *Ocimum basilicum*. *Clin Exp Pharmacol Physiol.* 2005; 32 (10): 811-6.
- [91] Kapewangolo P, Kandawa-Schulz M, Meyer D. Anti-HIV Activity of *Ocimum labiatum* Extract and Isolated Pheophytin-a. *Molecules.* 2017; 22 (11): 1763.
- [92] Gomez-Flores R, Verástegui-Rodríguez L, Quintanilla-Licea R, Tamez-Guerra P, Tamez-Guerra R, Rodríguez-Padilla C. *In vitro* rat lymphocyte proliferation induced by *Ocimum basilicum*, *Persea americana*, *Plantago virginica*, and *Rosa* spp. Extracts. *Journal of Medicinal Plants Research.* 2008; 2 (1): 005-010.
- [93] Eze CO, Nworu CS, Esimone CO, Okore VC. Immunomodulatory activities of methanol extract of the whole aerial part of *Phyllanthus niruri* L. *Journal of Pharmacognosy and Phytotherapy.* 2014; 6 (4): 41-46.
- [94] Utami PD, Ning R, Marsetyawan HS, Mubarika HS. Immune modulation properties of herbal plant leaves: *Phyllanthus niruri* aqueous extract on immune cells of tuberculosis patient - *in vitro* study. *Nat Prod Res.* 2018; 32 (4): 463-467.
- [95] Johnson JT, Ekpo GI, Ugwuoke JE. Immunomodulatory Potentials of Ethanolic Leaf Extract of *Phyllanthus amarus* in Wistar Rats. *The Pharmaceutical and Chemical Journal.* 2017; 4 (5): 83-88.

- [96] Wee CT, Indu BJ, Rishya M, Shamala DS. Evaluation of Antiviral Activities of Four Local Malaysian *Phyllanthus* Species against Herpes Simplex Viruses and Possible Antiviral Target. *Int J Med Sci.* 2013; 10 (13): 1817-1829.
- [97] Kumar S, Sharma S, Kumar D, Kumar K, Arya R. Immunostimulant activity of *Phyllanthusreticulatus*Poir: a useful plant for infectious tropical diseases. *Asian Pac. J. Trop. Dis.* 2004; 4 (Suppl. 1): S491-S495.
- [98] Deng Y, Chu J, Ren Y, Fan Z, Ji X, Mundy-Bosse B, Yuan S, Hughes T, Zhang J, Cheema B, Camardo AT, Xia Y, Wu LC, Wang LS, He X, Kinghorn AD, Li X, Caligiuri MA, Yu J. The natural product phyllanthusmin C enhances IFN-gamma production by human NK cells through upregulation of TLR-mediated NF-kappaB signaling. *J Immunol.* 2014; 193 (6): 2994-3002.
- [99] Nongluk S, Syuichi F, Kenji K, Hiroaki H, Takato O, Masato T, Yasuo S. Antiviral effects of *Psidiumguajava* Linn. (guava) tea on the growth of clinical isolated H1N1 viruses: its role in viral hemagglutination and neuraminidase inhibition. *Antiviral Res.* 2012; 94 (2): 139-46.
- [100] Trujillo-Correa AI, Quintero-Gil DC, Diaz-Castillo F, Quiñones W, Robledo SM, Martinez-Gutierrez M. *In vitro* and in silico anti-dengue activity of compounds obtained from *Psidiumguajava* through bioprospecting. *BMC Complementary and Alternative Medicine.* 2019; 19 (298).
- [101] Shrestha G, St Clair LL, O'Neil KL. The Immunostimulating Role of Lichen Polysaccharides: A Review. *PR.* 2015; 29 (3): 317-22.
- [102] Tariq J, Usman AA, Sana R, Rehman S, Riazuddin S. *In-vitro* antiviral activity of *Solanumnigrum* against Hepatitis C Virus. *Virology.* 2011; 8: 26.
- [103] Mohammed AH, Dhasarathan P, Dhanuskodi V. Evaluation of immunostimulant potential of *Solanumnigrum* L. using fish, *Etroplussuratensis* challenged with *Aphanomycesinvadens*. *International Journal of Pharma and Bio Sciences.* 2011; 2 (1): 429-437.4.
- [104] Li J, Li QW, Gao DW, Han ZS, Lu WZ. Antitumor and immunomodulating effects of polysaccharides isolated from *Solanumnigrum* L. *Phytother. Res.* 2009; 23: 1524-1530.
- [105] Soham S, Shyamasree G. *Tinosporacordifolia*: One plant, many roles. *AncSci Life.* 2012; 31 (4): 151-159.
- [106] Momoh M, Muhamed U, Agboke A, Akpabio E, Osonwa UE. Immunological effect of aqueous extract of *Vernoniaamygdalina* and a known immune booster called immunace® and their admixtures on HIV/AIDS clients: a comparative study. *Asian Pac J Trop Biomed.* 2012; 2: 81-184.
- [107] Onah I, Ebele O, Odimegwu D. Adjuvant effect of *Vernoniaamygdalina* leaf extract on host immune response to hepatitis B virus subunit vaccine. *Pharmazie.* 2019; 74 (3): 179-185.
- [108] Lidwina S, Jusak N, Pudji L, Sinansari R. Effect of African leaf (*Vernoniaamygdalina*) TO IL-6 and IL-10 level on *Staphylococcus aureus* infection. *Indonesian Journal of Tropical and Infectious Disease.* 2019; 7 (4): 69-74.
- [109] Sulochana K, Ginni J, Vaibhav K, Jaya PY, Samander K. Anti-viral activity of *Zingiberofficinale* (Ginger) ingredients against the Chikungunya virus. *VirusDisease.* 2020.
- [110] Chiang LC, Chiang W, Liu MC, Lin CC. *In vitro* antiviral activities of *Caesalpinia pulcherrima* and its related flavonoids. *Journal of Antimicrobial Chemotherapy.* 2003; 52 (2): 194-198.
- [111] Abd El-Wahab A, El-Adawi H, El-Demellawy M. *In vitro* study of the antiviral activity of *Zingiberofficinale*. *Planta Med.* 2009; 75-PF7.
- [112] CarrascoFR, Schmidt G, Lopez RA, Luiz SJ, Caparroz-Assef SM, Ciomar AB, Nakamura C. Immunomodulatory activity of *Zingiberofficinale* Roscoe, *Salvia officinalis* L. and *Syzygiumaromaticum* L. essential oils: evidence for humor- and cell-mediated responses. *J Pharm Pharmacol.* 2009; 61 (7): 961-7.
- [113] Masoud H, Mostafa SR. The effects of powdered ginger (*Zingiberofficinale*) on the haematological and immunological parameters of rainbow trout *Oncorhynchusmykiss*. *J. Med. Plant Herbal Ther. Res.* 2013; 1: 8-12.