



Utilization of Carbon Dioxide from Coal-Firing Flue Gas for Cultivation of *Spirulina platensis*

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To cite this article:

Oanh Thi Doan, Anh Kim Thi Bui, Kien Trung Hoang, Chuyen Hong Nguyen, Thom Thi Dang, Hong Diem Dang, Nguyet Thi Vu, Kim Dinh Dang. Utilization of Carbon Dioxide from Coal-Firing Flue Gas for Cultivation of *Spirulina platensis*. *American Journal of Environmental Protection*. Vol. 5, No. 6, 2016, pp. 152-156. doi: 10.11648/j.ajep.20160506.12

Received: October 14, 2016; **Accepted:** October 29, 2016; **Published:** November 18, 2016

Abstract: CO₂ emission from burning coal has been used as a carbon source for growing Cyanobacterium *Spirulina platensis* in order to minimize the cost of biomass production, and currently to carry out CO₂ bioremediation. This article presents the results of feeding *S. platensis* in laboratory conditions with 2 formulas including Pure CO₂ and Flue gas CO₂ upon using modified Zarrouk's medium with 1.6 g / L NaHCO₃ and 2g / L Na₂CO₃. Pure CO₂ with 1.2% concentrations taken from 99% vol of industrial CO₂ and CO₂ gas (1.2%) received from the flue gas through the Modular system of Exhausted Gas Treatment (MEGT). Growth of the Cyanobacterium using CO₂ - Flue gas is equivalent to CO₂ -Pure. On this basis, *S. platensis* has been cultivated outdoor in an 25 m² pond using CO₂ gas (1.2%) from the tunnel brick factory emissions after suitable cleaning. The experiment in an outdoor pond system of 25 m² indicated that the yield of biomass is of 10g/m²d with high-protein content (62.58 ± 2.34%) and fatty acids of high nutritional value (8.72 ± 0.14%), such as Omega - 6 and Omega - 3 reaching 14.74 ± 0.42% and 26.05 ± 0.64% of total fatty acid content, respectively. The quality of *Spirulina* cultured by CO₂ gas meets the requirements for functional foods according to Vietnam national food standards. The article also presents the results of biomass productivity and chemical composition of the Cyanobacterium in different culture conditions.

Keywords: CO₂, Carbon Source, Coal – Firing, Flue Gas, Cyanobacterium, *Spirulina platensis*

1. Introduction

CO₂ – anthropogenic carbon dioxide represents the most important greenhouse gases (GHGs) that contribute to approximately 77% of the global atmospheric temperature increase [1, 2]. The increase of CO₂ concentration in the atmosphere, mainly due to burning fossil fuels like coal, oil, gas and the forest destroying raises the deep concern about the climate change, and thereby, induces great challenges to the global sustainable development [3, 4].

Burning coal generates more carbon dioxide than any other

widely used fuel, including burning oil and gas that may harm the environment. Due to a rapid increase of the demand for the fossil fuels, there is a need for developing methods that allow continuous use of them through environmental friendly pathway along with reducing carbon dioxide emissions. In fact, there have been many efforts to reduce CO₂ emissions from burning fossil fuels. Overall, the current methods are focusing on CO₂ separation from emission sources and then trying to remove or capture it [5]. Some

other technologies such as chemical absorption and membrane separation, were also considered [6]. However, these methods can significantly reduce CO₂ concentration, they can not solve the problem of sustainable development [7].

Nowadays, in order to meet the demand for sustainable industrial development, it is highly desirable that exhaust emissions are treated thoroughly and sustainably through CO₂ recovery for use in photosynthesis. The recovery of CO₂ for microalgae culture is a novel pathway that has been studied and delivered in the reports. Lopes et al., 2008 indicated that as much as 40% of the carbon dioxide on the Earth can be absorbed by photosynthesis, in which microalgae or cyanobacteria make a great contribution with high species diversity and wide distribution in the ecological system [8]. So, photosynthesis by microalgae is an effective way to utilize CO₂ sources [9].

The selected culture strains have a considerable impact on biological fixation of CO₂ by level of temperature, SO_x, NO_x and CO₂ from flue gas [10]. Richmond mentioned that, cellular contents of *Spirulina platensis* were not changed by varying environmental conditions, compared with eukaryotic microalgae [11]. This alga is an excellent candidate for producing single cell protein due to its high protein content and nutritional value. Study on *S. platensis* due to potential of biomass production under high CO₂ concentration in flue gas is a good solution for CO₂ biofixation and for decreasing atmospheric CO₂ [12, 13].

Our objective is to assess the possibility of using CO₂ from coal combustion emissions for the growth of *Spirulina platensis*.

2. Materials and Methods

2.1. CO₂ Source

Pure CO₂ with 1.2% concentrations taken from 99% vol of industrial CO₂.

CO₂ received from the flue gas through the Modular system of Exhausted Gas Treatment (MEGT) described in the page [14].

2.2. Cyanobacterium and Cultivation Medium

Spirulina platensis strain, classified as *Arthrospira (Spirulina) platensis* used for the experiments, was supplied from the Collection of Microalgae and Cyanobacteria of Institute of Environmental Technology, Vietnam Academy of Science and Technology.

Culture medium: The medium for the microalgal growth is Zarrouk's medium modified by reducing NaHCO₃ to 1,36 g/L and by adding Na₂CO₃ to 2g/L [15].

2.3. Experimental Design

Laboratory Cultivation: *Spirulina platensis* cyanobacteria is cultivated in glass columns with a volume of 1 liter (inner diameter of 60 mm, height of 412 mm) which are maintained at a temperature of 27-32°C and illuminated by cold fluorescent light with intensity of 5,000 lux, and lighting time of 8 hours/day (Fig. 1.a). The liquid columns of Cyanobacterium are continuously bubbled with CO₂ - Flue gas (1.2 vol% CO₂) or CO₂ - Pure as control experiments (1.2 vol.% CO₂), at a rate of 50 L/min regulated by various valves. The pH of the medium is continuously controlled over time by pH equipment. Distilled water is added daily to eliminate evaporation effects during incubation.

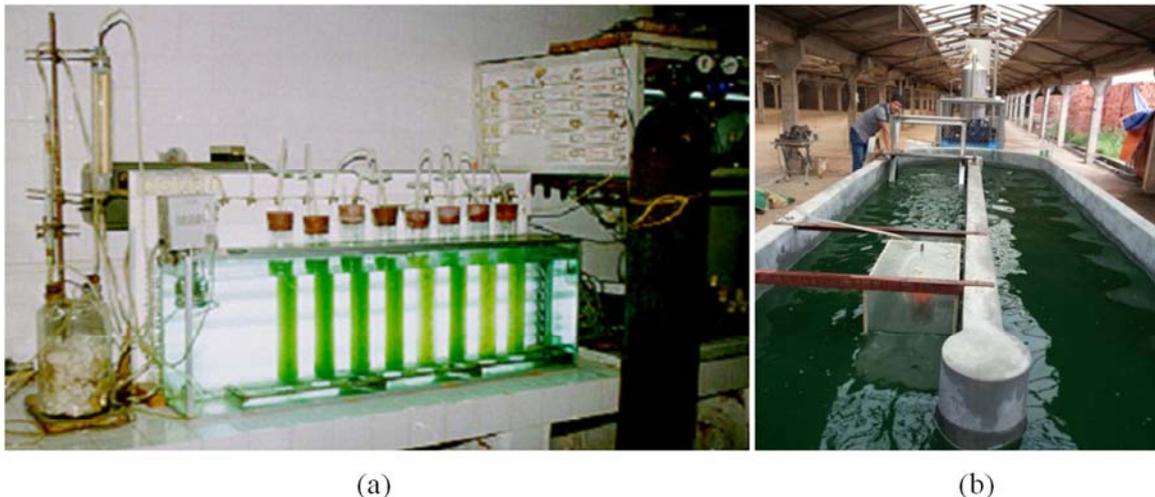


Fig. 1. *Spirulina platensis* culture system at laboratory conditions (a) and at outdoor conditions (b).

Spirulina platensis pond parameters have an area of 25 m² with culture depth of 0.25 m. This pond used 1.2% CO₂ from coal-fired flue gas which was cleaned via MEGT [14]. The experiments were carried out in 5 months (Fig. 1.b). The conditions for the Cyanobacterium culture: the average outdoor light intensity was about 25,000 lux, the temperature was in the range of 27 – 32°C.

The pH of the suspension is maintained at 8.5 - 9.5 and water is also added daily to eliminate evaporation effects. The pond was aerated by the paddle wheel system [16] in order to maintain moving speed of the suspension of about 18 cms⁻¹. The samples containing the *Spirulina* suspension were collected every two days for OD measurement at wavelength of 445nm using spectrophotometer UV-Vis 2450, Shimadzu,

Japan. Each month, the fresh biomass of the *Spirulina* was collected for quality analysis.

2.4. Sampling and Analysis

Samples were collected for biomass growth analysis (OD) and biomass quality analysis (lipids, fatty acids, total protein, fiber (%), carbohydrates, polysaccharide, ash, moisture and some important elements).

Lipids and fatty acids were analyzed according to the methods of Bligh and Dyer 1959 [17]. Total protein was determined by Kjeldahl method, multiplying by 6.25. Fiber, carbohydrates, ash, moisture were determined by the method of analysis AOAC 2000 [18]. Arsenic, Cd, Pb and Hg concentrations in *Spirulina* samples were measured using a Atomic Absorption Spectroscopy AA-6800, Shimadzu, Japan [19].

2.5. Data Analysis

All the data in mean and standard deviation were performed using Microsoft excel for Windows.

3. Results and Discussion

3.1. Growth of *S. platensis* in Two Formulas: Pure - CO₂ and Flue Gas - CO₂ at the Laboratory Scale

In the growth process, *Spirulina platensis* can use inorganic C sources under forms of CO₂, NaHCO₃ or Na₂CO₃ but the primary and most appropriate source is still HCO₃⁻ [13]. The supplementation of CO₂ to the algae culture

medium does not only provide C source but also control pH of the suspension. Fig. 2 presents the results of *Spirulina platensis* growth rate in 2 different formulas at the laboratory scale.

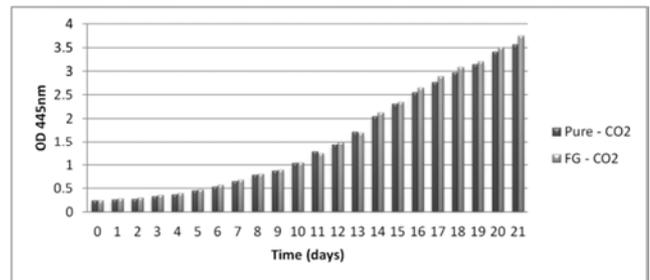


Fig. 2. The growth of *Spirulina platensis* with Pure CO₂ and Flue gas CO₂ at the laboratory scale.

After 21 days of experiment, *Spirulina platensis* increased biomass both in the 2 experimental formulas. Difference in *Spirulina platensis* biomass between two formulas (Pure CO₂ and FG CO₂) is negligible. However, in the last week of the experiment, the growth of *Spirulina platensis* in FG- CO₂ formula was slightly better than in Pure-CO₂ formula. The achieved results may be explained that there is a small amount of NO_x as nutrient for algae besides CO₂ in the coal burning emissions [20].

In addition to assessing efficiency of the above 2 sources of CO₂ on the growth of *S. platensis*, we also analyzed the nutritional composition of the biomass of this Cyanobacterium (Table 1).

Table 1. Chemical composition of fresh biomass of *S. platensis* cultured in different experimental formulas (per 100 g dry weight \pm 5.43 g).

Parameters	Unit	Pure - CO ₂	FG - CO ₂	Parameters	Unit	Pure - CO ₂	FG - CO ₂
Moisture	g	2.51 \pm 0.04	2.46 \pm 0.03	Lead (Pb)	ppm	0.54 \pm 0.02	0.57 \pm 0.03
Protein	g	61.32 \pm 1.48	61.21 \pm 1.34	Cadmium (Cd)	ppm	< 0.01	< 0.01
Fat (Lipids)	g	8.63 \pm 0.19	8.68 \pm 0.12	Arsenic (As)	ppm	0.13 \pm 0.01	0.06 \pm 0.01
Fibre	g	0.4 \pm 0.05	0.39 \pm 0.06	Mercury (Hg)	ppm	< 0.01	< 0.01
Ash	g	8.52 \pm 0.35	8.64 \pm 0.27	Others	g	18.62 \pm 1.22	18.62 \pm 1.08
Carotenoids	mg	121 \pm 5.26	149 \pm 6.62				

The research results presented in Table 1 show that there is no difference in chemical composition of the biomass between two formulas and the use of CO₂ from coal-fired emissions for *Spirulina platensis* cultivation has been proved to be advantageous, and could be applied in large scale.

3.2. Growth and Productivity of *Spirulina platensis* in Culture Conditions at Dan Phuong, Hanoi

Optical density (OD) measurement for *Spirulina platensis* growth was applied in our study. The OD_{445nm} and dry biomass of the *Spirulina platensis* were determined every two days. The results in Fig. 3 demonstrated the variation of OD_{445nm} of the culture suspension. On the first two days *S. platensis* grew slowly, the OD_{445nm} value increased from 0.21 to 0.35. After ten days, *S. platensis* grew rapidly from 0.34 - 0.35 to 1.09 - 1.11. The highest level of OD_{445nm} in this experimental process reached up to 1.67 - 1.73 when the

biomass harvest was performed for maintaining the algal OD relatively constant.

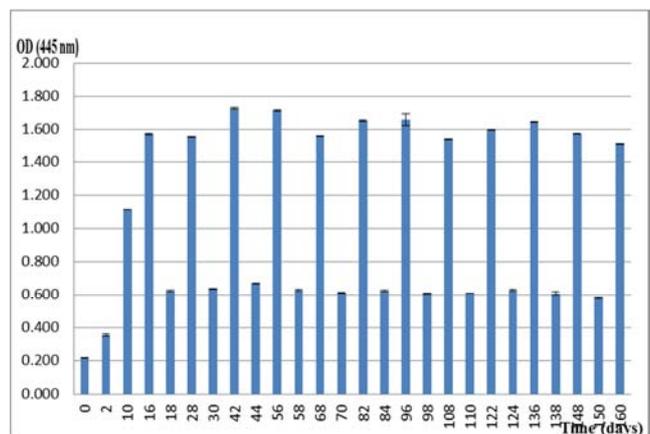


Fig. 3. *Spirulina platensis* growth at outdoor conditions.

Carbon dioxide is well adsorbed inside the *S. platensis* culture medium with pH > 8.5. During the photosynthesis, alkaline medium is normally created through the metabolic processes by phototrophic microorganisms participating in the transport of hydroxide ion (OH⁻) outwards its cell through catalytic reaction by carbohydrate anhydrase. As a result, the medium with phototrophic organisms as *Spirulina platensis* displays a strong alkaline property that helps them adsorb CO₂ with high efficiency [21]. Therefore, there have been a lot of studies taking into account the microalgae using CO₂ for nutritive biomass production. Cheng *et al.*, (2006) has cultured *Chlorella vulgaris* in photobioreactor presenting that its growth rate is good in the medium with 1% CO₂ [22]. *Aphanothece microscopica* Nägeli (RSMAN92) was cultured in tubular photobioreactors with different concentration of the carbon dioxide (3, 15, 25, 50 and 62%), light intensity (960, 3000, 6000, 9000 and 11000 lux), and temperature (21.5, 25, 30, 35 and 38.5°C) in order to determine the optimum condition, the highest CO₂ absorption processes for this microalgae strain [8]. In the study of Sydney *et al.* (2010), *Botryococcus braunii* presented the highest CO₂

fixation rate, followed by *Spirulina platensis*, *Dunaliella tertiolecta*, *Chlorella vulgaris* (as 496.98, 318.61, 272.4, and 251.64 mg l⁻¹ d⁻¹, respectively) [23].

In this research, flue-gas emissions from coal combustion contained CO₂ with the amount around of 1.2%. Using such a gas source for the algae production pond with the aeration time of 6 to 8 hr d⁻¹ made the pH of their medium not to raise sharply, and keeps the pH unchanged in the range of 8.5-9.6. Additionally, using a modified Zarrouk's medium (the content of bicarbonate reduced to 1.6 gl⁻¹ NaHCO₃ and adding 2 gl⁻¹ carbonate - Na₂CO₃) in which CO₂ extracted from coal-fired gas gave good results in *Spirulina* growth during the 180-day experiment.

For assessing the growth and productivity of *S. platensis* using CO₂ in the outdoor conditions, the analysis of chemical composition in the Cyanobacterium biomass was also carried out (Table 2 and 3). The obtained results presented in Table 2 indicated that *S. platensis* was rich in protein, reaching up to 62.69% dry weight while the lipid content did not exceed 9%.

Table 2. Biomass quality of *S. platensis* cultivated outdoor after spray drying (per 100 g dry weight ± 5.02g).

Parameters	Unit	CO ₂ from flue gas via MEGT	Parameters	Unit	CO ₂ from flue gas via MEGT
Moisture	g	2.39 ± 0.04	Lead (Pb)	ppm	0.14 ± 0.01
Protein	g	62.58 ± 2.34	Cadmium (Cd)	ppm	< 0.01
Fat (Lipids)	g	8.72 ± 0.14	Arsenic (As)	ppm	0.04 ± 0.01
Fibre	g	0.43 ± 0.03	Mercury (Hg)	ppm	< 0.01
Ash	g	9.83 ± 0.06	Others	g	16.05 ± 0.97
Carotenoids	mg	44 ± 3.44			

Moreover, the *Spirulina* also contained fatty acids having high nutritional value, such as Omega - 6 and Omega - 3 which reached 14.74% and 26.05% of total fatty acid content, respectively (Table 3).

Table 3. Composition of fatty acids in biomass after spray drying.

Fatty acids	Scientific name	Quantity (%)
14:0	Pentadecanoic acid	ND
16:0	Hexadecanoic acid	45.48 ± 1.24
16:1n-7	9-Hexadecenoic acid	4.43 ± 0.12
17:0	Heptadecanoic acid	ND
17:1n-5	Heptadecenoic acid	ND
18:0	Octadecanoic acid	ND
18:1n-6	Octadecenoic acid	3.59 ± 0.08
18:3n-3	9,12,15-octadecatrienoic acid	26.05 ± 0.64
18:3n-6	6,9,12-octadecatrienoic acid	14.74 ± 0.42
20:0	Eicosanoic acid	5.71 ± 0.09
20:3n-6	11,14,17-eicosatrienoic acid	ND
20:4n-6	5,8,11,14-eicosatetraenoic acid	ND

ND: non detection

As shown in Fig. 3, during the experimental production of the *Spirulina* at the outdoor conditions, the Cyanobacterium was harvested when OD₄₄₅ values achieved about 1.6 with the productivity of about 10g/m²d. This showed that the using CO₂ from coal-fired flue gas for culturing *Spirulina platensis* is feasible for mass culture.

The quality of *Spirulina* cultured by CO₂ gas from coal-

fired flue gas of the Tuynel Brick Factory is good and equivalent to that of Siam Algae Company (SAC) assessed by Japan Food Researcher Laboratories [24]. Heavy metal concentrations, including Pb, Cd, As, Hg, and the others (Table 2), of the *Spirulina* remained within acceptable limits for functional foods according to the Decision No. 46/2007/QĐ-BYT and VNTR 8-2:2011/BYT (Vietnam) [25, 26]. This is an important basis for using *Spirulina* as a nutritive food or functional food source for humans. In the context of global climate change, *Spirulina platensis* not only makes a positive contribution to reducing greenhouse gases - CO₂ but could be also good biomass for different purposes.

4. Conclusion

At the laboratory scale, biomass growth and quality of *Spirulina platensis* in 2 formulas (Pure CO₂ and Flue gas CO₂) are equivalent. The experiment in an outdoor pond system of 25 m² indicated that the yield of biomass is of 10g/m²d with high-protein content of 62.58 ± 2.34% and fatty acids of high nutritional value (8.72 ± 0.14%), such as Omega - 6 and Omega - 3 reaching 14.74% ± 0.42 and 26.05 ± 0.64% of total fatty acid content, respectively. The obtained results allowed evaluating the potential of using CO₂ from coal combustion emissions for *S. platensis* culture with cost effective way for carbon sources and also for environment protection.

Nomenclature

GHGs: Greenhouse Gases

MEGT: Modular system of Exhausted Gas Treatment

OD: Optical Density

VNNTR: Viet Nam National technical regulation

Acknowledgements

This work was financially supported by The National Project KC08.08/11-15, Ministry of Science and Technology (MOST), Vietnam.

References

- [1] IPCC - The United Nations Intergovernmental Panel on Climate Change: Climate Change 2007 Mitigation. (2007) Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change [B. Metz, O. R. Davidson, P. R. Bosch, R. Dave, L. A. Meyer (eds)], Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 841 pp.
- [2] Juan, C. L., Guillermo, Q., Theo, S. O. S., José, M. E., Raquel, L., Raúl, M. (2013) Biotechnologies for greenhouse gases (CH₄, N₂O, and CO₂) abatement: state of the art and challenges. *Appl Microbiol Biotechnol* 97: 2277–2303.
- [3] Maroto-Valer, M. M., Song, C., Soong, Y., (Eds). (2002) Environmental Challenges and Greenhouse Gas Control for Fossil Fuel Utilization in the 21st Century. Kluwer Academic/Plenum Publishers, New York, 447 pp.
- [4] Song, C., Gaffney, A. M., Fujimoto, K., (Eds). (2002) CO₂ Conversion and Utilization. American Chemical Society (ACS), Washington DC, ACS Symp Series Vol 809 448 pp.
- [5] Iglesias-Rodríguez, M. D., Halloran, P. R., Rickaby, R. E. M., Hall, I. R., Colmenero-Hidalgo, E., Gittins, J. R., Green, D. R. H., Tyrrell, T., Gibbs, S. J., Dassow, P. V., Rehm, E., Armbrust, E. V., Boessenkool, K. P. (2008) Phytoplankton Calcification in a High-CO₂ World. *Science* 320 (5874): 336–340.
- [6] Aiba, S., Ogawa, T. (1997) Assessment of growth yield of a blue-green alga: *Spirulina platensis*, in axenic and continuous culture. *J of General Microbiology* 102: 179–182.
- [7] Kim, D. D., T. V. Tua, N. T. Cu, D. T. Anh, D. T. Thom, H. T. Kien, L. T. T. Thuy, T. V. Nguyet, M. T. Chinh, & N. V. Vuong. (2011) Utilization of CO₂ captured from the coal-fired fuel gas for growing *Spirulina platensis* SP4. *Journal of Science and Technology* 49 (4): 65 – 72 ISSN 0866 708X.
- [8] Lopes, E. J., Scoparo, C. H. G., Franco, T. T. (2008) Rates of CO₂ removal by *Aphanothece microscopica Nageli* in tubular photobioreactors. *Chemical Engineering and Processing* 47: 1365–1373.
- [9] Uday, B. S., Ahluwalia, A. S. (2013) Microalgae: a promising tool for carbon sequestration. *Mitig Adapt Strateg Glob Change* 18: 73–95.
- [10] Kumar, K., Dasgupta, C. N., Nayak, B., Lindblad, P., Das, D. (2011) Development of suitable photobioreactor for CO₂ sequestration addressing global warming using green algae and cyanobacteria. *Bioresour Technol* 102: 4945–4953.
- [11] Richmond, A. (2013) Handbook of microalgal culture: biotechnology and applied phycology. 3rd ed. Oxford: Blackwell Science Ltd.
- [12] Seyedmahdi, H., Saeed, A., Mohamad, S. H., Fatemeh, M. (2014) Growth response of *Spirulina platensis* PCC9108 to elevated CO₂ levels and flue gas. *Biological Journal of Microorganism*, 29- 36.
- [13] Song-Gun, K., Chan-Sun, P., Yong-Ha, P. (2004) Effect of CO₂ Concentration on Growth and Photosynthesis of *Spirulina platensis*. *Studies in Surface Science and Catalysis*, 153: 295–298.
- [14] Nguyet M. T. T., et al., (2013) Application studies on catalytic nano materials for removal of hazardous gases. *Journal of Catalysis and adsorbent*, 3: 136–142.
- [15] Aiba S., Ogawa T., 1997. Assessment of growth yield of a blue-green alga: *Spirulina platensis*, in axenic and continuous culture. *J. Gen. Microbiology*, 10: 179–182.
- [16] Masoji'dek, J., Torzillo, G. (2014) Mass Cultivation of Freshwater Microalgae. Reference Module in Earth Systems and Environmental Sciences, 13 pp.
- [17] Bligh, E. G., Dyer, W. J. (1959) A rapid method for total lipid extraction and purification. *Can J Biochem Physiol* 37: 911–917.
- [18] Official Methods of Analysis. (2000) 17th Ed., AOAC INTERNATIONAL, Gaithersburg, MD.
- [19] Horwitz, W. (2000) Official method of analysis of AOAC International. Published by AOAC International Suite 500, 481 North Frederick Avenue, Gaithersburg, Maryland 20877-2471, USA.
- [20] Negoro, M., Shioji, N., Miyamoto, K. and Miura, Y. (1991) Growth of microalgae in high CO₂ gas and effects of SO_x and NO_x. *Appl. Biochem. Biotechnol.*, 28/29, 877–886.
- [21] Simona, A., Carlo, S., Alessandra, L., Adriana, D. B. (2013) *Spirulina platensis* Culture with Flue Gas Feeding as a Cyanobacteria-Based Carbon Sequestration Option. *Chem Eng Technol* 36 (1) 91–97.
- [22] Cheng, L., Zhang, L., Chen, H., Gao, C. (2006) Carbon dioxide removal from air by microalgae cultured in a membrane-photobioreactor. *Separation and Purification Technology* 50: 324–329.
- [23] Sydney, E. B., Sturm, W., Carvalho, J. C., Soccol, V. T., Larroche, C., Pandey, A., Soccol, C. R. (2010) Potential carbon dioxide fixation by industrially important microalgae. *Bioresource Technology* 101: 5892–5896.
- [24] Hidenori, S. (2004) Mass production of *Spirulina*, an edible microalga. *Asian Pacific Phycology in the 21 st Century: Próspects and Challenges Developments in Hydrobiology* 173 39 – 44.
- [25] Decision No. 46/2007/QĐ-BYT dated December 19, 2007 of the Viet Nam Ministry of Health on Promulgation regulation of maximum level of biological and chemical pollution in food.
- [26] VNNTR 8-2: 2011/BYT of the Viet Nam Ministry of Health on National technical regulation on the limits of heavy metals contamination in food.