

Synthesis, Characterization and in vitro Antimicrobial studies of Schiff-bases derived from Acetyl acetone and Amino acids and their Oxovanadium(IV) complexes

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Abstract: A series of novel amino acid derived Schiff-bases and their Oxovanadium(IV) complexes were synthesized and well characterized by elemental analyses, spectral studies, conductivity and magnetic measurements. Physical and analytical data suggest that the Schiff bases act as tridentate ligands towards metal ion via azomethine-N, deprotonated-O of carboxylic group and enolic-O group of acetylacetone. All the complexes have lower molar conductance values, indicating their non-electrolytic nature. The synthesized ligands, along with their metal complexes were screened for their *in-vitro* antibacterial activity against two Gram-negative (*Escherichia coli*, *Salmonella typhi*) and two Gram-positive (*Bacillus subtilis*, *Staphylococcus aureus*) bacterial strains and for in vitro antifungal activity against *Trichophyton longifusus*, *Candida albicans*, *Aspergillus flavus*, and *Candida glabrata* species. The results of these studies revealed that all the compounds and their metal complexes showed significant antibacterial and antifungal potency. Brine shrimp bioassay was also carried out for *in vitro* cytotoxic properties against *Artemia salina*.

Keywords: Schiff-bases, Amino Acid, Antibacterial, Antifungal, Cytotoxicity

1. Introduction

Schiff base are an important class of ligands in coordination chemistry due to not only their useful physical and chemical properties and large number of reactions they undergo but also have very wide use in industry and due to their interesting pharmacological as well as physiological activities.[1,2] They are also important intermediate in the enzymatic reactions involving interaction of an enzyme with an amino or a carbonyl group of the substrate.[3] It is possible to introduce different substituents into the existing skeleton of the molecule, hence enabling the designing of compounds with the suitable structural, electronic and biological properties.[4] In recent years, Schiff base complexes of amino acid have received considerable attention because of their physiological and pharmacological activities.[5,6] In metal coordination chemistry Schiff bases continue to occupy an important position as ligand ions due to the incorporation of transition metals into Schiff bases. This results in an increase in

biological activity of the ligand and decrease in the cytotoxic effects of both the metal ion and ligand on the host.[7] Metal ions play a catalytic role in reactions of Schiff bases. The development in the field of bioinorganic chemistry has increased the interest in Schiff base complexes, since it has been recognized that many of these complexes may serve as models for biologically important species. It has also been reported that metal complexes of amino acid Schiff bases possess anti-carcinogenic activities.[8] The coordination behavior of Schiff bases with Vanadium is of great interest because of the presence of Vanadium in biological systems as trace element. The geometry of vanadyl complexes is highly dependent on the chelating properties of ligands.[9,10] It has been observed that Schiff bases form stable complexes with oxovanadium(IV) salts with five or six coordinated state of V(IV).[11] A square pyramidal or distorted trigonal bipyramidal geometry is reported for five coordinated complexes[12] while distorted octahedral structure is observed for the six coordinated ones.[13] In past few years attentions have been focused on the synthesis of low

toxicity bacteriostatic agents and it have been observed that Vanadyl ion is less toxic than vanadate ion. Keeping in view the importance of Schiff base oxovanadium(IV) complexes, in present studies, we have prepared four ligands (L_1 - L_4) by the condensation reaction between amino acids (Tyrosin, Threonin, Lysin, Arginin) and acetylacetone with this hope that it may provide us valuable theoretical information for exploring metal-based anti-microbial and carcinogenic agents with high efficiency and low toxicity. The synthesized amino acid-derived schiff bases (L_1 - L_4) have been exposed to act as tri-dentate towards vanadyl ion through the azomethine-N, carboxylat-O and enolic-O groups forming 5 and 6 membered stable chelate ring system.

2. Experimental

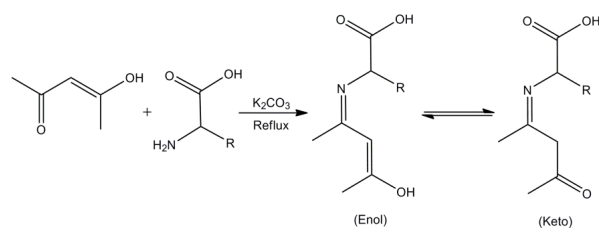
2.1. Materials and Physical Measurements

All the reagents and solvents were purchased from Sigma-Aldrich and they were used as received. Reactions were monitored by thin layer chromatography (plates coated with 0.2 mm Merck 60 F254 silica gel) and were visualized by UV irradiation (254 nm). Elemental analyses were carried out with a LECO-CHNS-9320 model. ^1H and ^{13}C -NMR spectra of compounds were recorded with a Bruker Spectrospin Avance DPX-400 using TMS as internal standard and d_6 DMSO as solvent. Infrared spectra of compounds were recorded on a Philips Analytical PU 9800 FTIR spectrophotometer. The melting points of compounds were determined with a Gallenkamp melting point apparatus. UV/visible absorption spectra were recorded using a Shimadzu UV-1700 spectrophotometer at room temperature. Conductance was recorded by pre-calibrated cyber scan 500 conductivity meter. Electron impact mass spectra (EIMS) were recorded on a JEOL MS Route instrument. Thermogravimetric analysis (TGA) was carried out under constant nitrogen flow at a heating rate of $15^\circ\text{C min}^{-1}$, using a Mettler Toledo TGA/SDTA 851 balance. The heating scans were performed on 3-5 mg of sample, in the temperature range 25 - 900°C . In vitro antibacterial, antifungal and cytotoxic properties were studied at HEJ Research Institute of Chemistry, International Center for Chemical Sciences, University of Karachi, Pakistan.

2.2. Synthesis and Characterization of Schiff Bases

To a stirred solution of respective amino acid (20 mmol in 20 ml water) was added acetylacetone (20 mmol in 10 ml ethanol). To this solution K_2CO_3 (0.1% in ethanol) was added to adjust the pH of the solution at 7-8. The resulting reaction mixture was refluxed on water bath for 3 h. The completion of the reaction was monitored by TLC. After completion of the reaction, the volume of the mixture was reduced to half in vacuo. On cooling a solid product was

formed, filtered, washed with ethanol, then with ether, and dried. (Scheme 1).

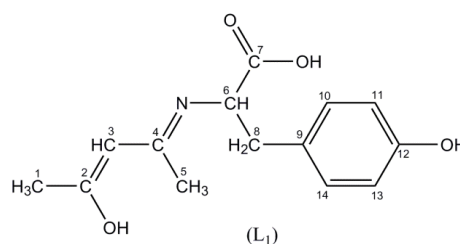


Scheme 1. Synthesis of Schiff bases.

2.3. Compound 1, (L_1)

(*R*)-2-((4-Hydroxypent-3-En-2-Ylidene) Amino)-3-(4-Hydroxyphenyl) Propanoic Acid

Yellow solid; yield 70%; m.p: 210°C ; Anal. Calcd. For $\text{C}_{14}\text{H}_{17}\text{NO}_4$: C, 63.64; H, 6.62; N, 5.28%. Found: C, 63.87; H, 6.51; N, 5.32%; IR (KBr, cm^{-1}): 3436 (-OH stretching), 3055 (-CH stretching of aromatic ring), 2962 (-CH stretching of CH_3), 2922 (-CH stretching of aliphatic group), 1708 (-C=O stretching), 1625 (C=C stretchig), 1610 (C=N stretching), 1346 (C-N stretching), 1236 (C-O stretching of phenolic group); ^1H -NMR (400 MHz, DMSO-d_6 , δ / ppm): 12.2 (1H, s, OH), 11.3 (1H, s, COOH), 8.7 (1H, s, OH, Phenolic), 7.2 (2H, d, Aromatic, $J = 1.5$ Hz), 6.8 (2H, d, Aromatic, $J = 3.8$ Hz), 5.1 (1H, s, HC=C), 2.9 (1H, m, N-CH-C), 2.7 (2H, d, Aliphatic CH_2 , $J = 7.5$ Hz), 2.2 (3H, s, CH_3), 2.1 (3H, s, N=C- CH_3); ^{13}C NMR (400 MHz, DMSO-d_6 , δ / ppm): 180.2 (C7, COO), 178.3 (C2, COH), 162.1 (C4, C=N), 157.6 (C12, Aromatic), 128.2 (C9, Aromatic), 122.5 (C10 and C14 Aromatic), 112.6 C11 and C13, Aromatic), 80.2 (C3, CH), 71.1 (C6, CH), 40.3 (C8, Aliphatic CH_2), 27.6 (C1, CH_3), 14.5 (C5, CH_3); MS (ESI) $[\text{M}]^+$: 263.



2-((*E*)-(Z)-4-hydroxypent-3-en-2-ylidene)amino)-3-(4-hydroxyphenyl)propanoic acid

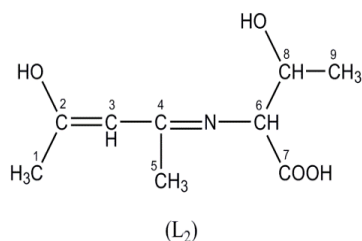
Figure 1. Structure of the ligand (L_1)

2.4. Compound 2, (L_2)

(2*S*)-3-Hydroxy-2-((4-Hydroxypent-3-En-2-Ylidene) Amino) Butanoic Acid

Yellow solid; yield 68%; m.p: 208°C ; Anal. Calcd. For $\text{C}_9\text{H}_{15}\text{NO}_4$: C, 53.72; H, 7.51; N, 6.96. Found: C, 53.69; H, 7.53; N, 6.92; IR (KBr, cm^{-1}): 3446 (-OH stretching), 2954 (-CH stretching of CH_3), 2896 (-CH stretching of aliphatic CH), 1714 (-C=O stretching), 1632 (C=C stretchig), 1614 (C=N stretching), 1342 (C-N stretching), 1104 (C-O stretching of alcoholic group); ^1H -NMR (400 MHz, DMSO-d_6 , δ / ppm): 12.8 (1H, s, C=C-OH), 11.2 (1H, s,

COOH), 5.2 (1H, s, HC=C), 4.1 (1H, s, C-OH), 3.8 (1H, m, CH), 2.4 (1H, d, N-CH), 2.3 (3H, s, C=C-CH₃), 2.1 (3H, s, N=C-CH₃), 1.4 (3H, d, CH₃, J = 7.4 Hz); ¹³C NMR (400 MHz, DMSO-d₆, δ / ppm): 175.5 (C7, COO), 172.2 (C2, C-OH), 162.4 (C4, C=N), 80.1 (C3, HC=C), 68.5 (C8, HC-OH), 64.3 (C6, N-CH), 20.6 (C9, CH₃), 17.5 (C1, CH₃), 14.5 (C5, CH₃); MS (ESI) [M]⁺: 201.



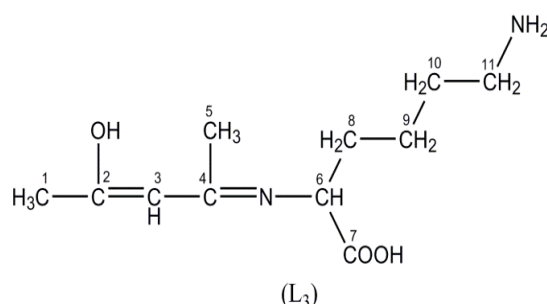
(2S)-3-hydroxy-2-((4-hydroxypent-3-en-2-ylidene)amino)butanoic acid

Figure 2. Structure of the ligand (L₂)

2.5. Compound 3, (L₃)

(S)-6-Amino-2-((4-Hydroxypent-3-En-2-Ylidene) Amino) Hexanoic Acid

Yellow solid; yield 74%; m.p: 194 °C; Anal. Calcd. For C₁₁H₂₀N₂O₃: C, 57.87; H, 8.83; N, 12.27. Found: C, 57.83; H, 8.76; N, 12.35; IR (KBr, cm⁻¹): 3438 (-OH stretching), 3392 (NH stretching), 2962 (-CH stretching of CH₃), 2932 (-CH stretching of CH₂), 2892 (-CH stretching of aliphatic CH), 1720 (-C=O stretching), 1625 (C=C stretchig), 1606 (C=N stretching), 1586 (NH bending), 1337 (C-N stretching), 1102 (C-O stretching of alcoholic group); ¹H-NMR (400 MHz, DMSO-d₆, δ / ppm): 12.4 (1H, s, C-OH), 11.2 (1H, s, COOH), 5.3 (2H, m, NH₂), 5.1 (1H, s, HC=C), 2.8 (2H, m, N-CH₂), 2.5 (1H, m, N-CH), 2.3 (3H, s, CH₃), 2.1 (3H, s, N=C-CH₃), 1.8 (2H, m, CH₂), 1.6 (2H, m, CH₂), 1.3 (2H, m, CH₂); ¹³C NMR (400 MHz, DMSO-d₆, δ / ppm): 180.2 (C7, COO), 174.2 (C2, C=CH), 168.2 (C4, C=N), 82.4 (C3, CH=C), 65.2 (C6, N-CH), 45.2 (C11, CH₂-NH₂), 36.2 (C10, CH₂), 34.4 (C8, CH₂), 24.2 (C9, CH₂), 19.6 (C1, CH₃), 14.2 (C5, CH₃); MS (ESI) [M]⁺: 228.



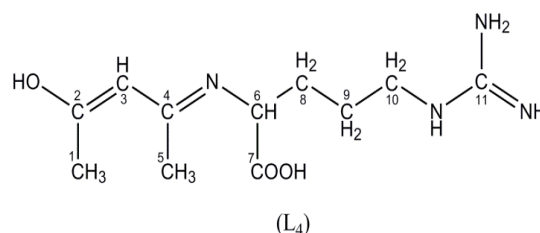
(S)-6-amino-2-((4-hydroxypent-3-en-2-ylidene)amino)hexanoic acid

Figure 3. Structure of the ligand (L₃)

2.6. Compound 4, (L₄)

5-Guanidino-2-((E)-((E)-4-Hydroxypent-3-En-2-Ylide ne) Amino) Pentanoic Acid

Yellow solid; yield 72%; m.p: 185 °C; Anal. Calcd. For C₁₁H₂₀N₄O₃: C, 51.55; H, 7.87; N, 21.86. Found: C, 51.47; H, 7.68; N, 21.92; IR (KBr, cm⁻¹): 3442 (-OH stretching), 3375 (NH stretching), 2970 (-CH stretching of CH₃), 2940 (-CH stretching of CH₂), 2898 (-CH stretching of aliphatic CH), 1714 (-C=O stretching), 1632 (C=C stretchig), 1610 (C=N stretching), 1595 (NH bending of primary amine), 1565 (NH bending of secondary amine), 1337 (C-N stretching), 1298 (C-N stretching of secondary amine), 1276 (C-N stretching of primary amine), 1095 (C-O stretching of alcoholic group); ¹H-NMR (400 MHz, DMSO-d₆, δ / ppm): 12.8 (1H, s, C-OH), 11.2 (1H, s, COOH), 8.4 (2H, s, NH₂), 5.2 (1H, s, HC=C), 4.8 (1H, m, HN=C), 3.4 (1H, m, NH), 2.7 (2H, m, CH₂-NH), 2.5 (1H, m, HC-N), 2.3 (3H, s, CH₃), 2.1 (3H, s, N=C-CH₃), 1.8 (2H, m, CH₂), 1.5 (2H, m, CH₂); ¹³C NMR (400 MHz, DMSO-d₆, δ / ppm): 179.2 (C7, COO), 176.2 (C2, C-OH), 162.4 (C4, C=N), 156.5 (C11, C=NH), 85.2 (C3, HC=C), 70.2 (C6, HC-N), 42.6 (C10, HN-CH₂), 38.4 (C8, CH₂), 26.6 (C9, CH₂), 18.6 (C1, CH₃), 14.2 (C5, CH₃); MS (ESI) [M]⁺: 256.



5-guanidino-2-((E)-((E)-4-hydroxypent-3-en-2-ylidene)amino)pentanoic acid

Figure 4. Structure of the ligand (L₄)

2.7. Synthesis of Oxovanadium(IV) Complexes

To a hot magnetically stirred ethanol solution of Schiff bases (L₁-L₄, 0.1 mole), an ethanolic solution of metal salt (0.1 mole) was added. The mixture was then refluxed for 2-12 h. The completion of reaction was monitored by TLC. The solution obtained was cooled at room temperature, precipitates appeared were filtered and washed with ethanol and dried. (Scheme 2).



where;

M = VO

L = L₁, L₂, L₃, L₄

Scheme 2. Synthesis of oxovanadium(IV) complexes.

2.8. Antibacterial Activity

The *in-vitro* antibacterial activity of Schiff bases (L₁-L₄) and their oxovanadium complexes (C₁-C₄) was assayed against two Gram-negative (*Escherichia coli*, *Salmonella typhi*) and two Gram-positive (*Staphylococcus aureus*, *Bacillus subtilis*) bacterial strains by the reported method.[14] The stock solution (1 mg/ml) of the test chemical was prepared by dissolving 10 mg of the test compound in 10 ml of Dimethyl sulfoxide (DMSO) solvent. The stock solution was suitably diluted with sterilized distilled water to get dilution of 100, 50 and 25 mgml⁻¹. Control for each dilution was prepared by diluting 10 ml of solvent instead of stock solution with sterilized distilled water. The wells (6 mm in diameter) were dug in the agar media with the help of a sterile metallic borer. Two to eight hours old bacterial inocula containing approximately 10⁴-10⁶ colony forming units (CFU/mL) were spread on the surface of the nutrient agar with the help of a sterile cotton swab. The prepared concentrations of the test sample were introduced in the respective wells. Other wells supplemented with DMSO and reference antibacterial drug, Imipenem, served as negative and positive controls, respectively. The plates were incubated immediately at 37 °C for 24 h. Activity was determined by measuring the diameter of zones showing complete inhibition (mm). In order to clarify any effect of DMSO in the biological screening, separate studies were carried out with the solutions alone of DMSO and they showed no activity against any bacterial strains.

2.9. Antifungal Activity

All the compounds (L₁-L₄) and their oxovanadium complexes (C₁-C₄) were studied against four fungal cultures (*Trichophyton longifusus*, *Candida albicans*, *Aspergillus flavus*, and *Candida glabrata*) for Antifungal activities. Sabouraud dextrose agar (Oxoid, Hampshire, England) was seeded with 10⁵ cfu ml⁻¹ fungal spore suspensions and transferred to petri plates. The stock solution of test chemical was prepared and diluted to 100, 50 and 25 mgml⁻¹. Discs soaked in 20 ml of prepared concentrations of all compounds were placed at different positions on the agar surface. The plates were incubated at 32 °C. The percentage inhibition was calculated after seven days and compared with standard drugs Miconazole.[15]

2.10. In Vitro Cytotoxicity

The synthesized compounds and their oxovanadium(IV) complexes were screened for their cytotoxicity (brine shrimp bioassay) by using the protocol of Meyer et al. [16] Brine shrimp (*Artemia salina* leach) eggs were hatched in a shallow rectangular plastic dish (22 x 32 cm) filled with artificial seawater, which was prepared with a commercial salt mixture and double distilled water. An unequal partition was made in the plastic dish with the help of a

perforated device. Approximately 50 mg of eggs were sprinkled into the large compartment, which was darkened while the minor compartment was open to ordinary light. After two days nauplii were collected by a pipette from the lighted side. A sample of the test compound was prepared by dissolving 20 mg of each compound in 2 ml of DMSO. From this stock solution 100, 50 and 25 mgml⁻¹ were transferred to nine vials (three for each dilutions were used for each test sample and LD₅₀ is the mean of three values) and one vial was kept as control having 2 ml of DMSO only. The solvent was allowed to evaporate overnight. After two days, when shrimp larvae were ready, 1 ml of seawater and 10 shrimps were added to each vial (30 shrimps/dilution) and the volume was adjusted with seawater to 5 ml per vial. After 24 h the number of survivors was counted. Data were analyzed by a Finney computer program to determine the LD₅₀ values.[17]

3. Results and Discussion

3.1. IR Spectra

In order to study the bonding mode of ligand to metal in the complexes, IR spectrum of free ligands was compared with the complexes. The structurally significant IR band for ligands and their complexes have been reported in the Experimental and Table I respectively. Acetylacetone is capable of exhibiting keto-enol tautomerism and react with metal cations to form metal complexes. The IR spectra of all the ligands show the absence of bands at 3245 and 1745 cm⁻¹ due to $\nu(\text{HN}_2)$ group of amino acids and $\nu(\text{C=O})$ of acetylacetone.[18] Instead, a new prominent band at 1606-1614 cm⁻¹ due to azomethine $\nu(\text{C=N})$ linkage appeared in all the ligands indicating [19] that condensation between ketone moiety of acetylacetone and that of amino group of amino acid has taken place resulting into the formation of the desired ligands (L₁-L₄). IR spectra of the oxovanadium(IV) complexes showed[20] a major shift to lower wave numbers by 15-20 cm⁻¹ in azomethine $\nu(\text{C=N})$ suggesting involvement of the azomethine-N with the metal(II) ion. This is further supported by the appearance of new bands at 450-486 cm⁻¹ due to $\nu(\text{M-N})$ bond.[21] Also, disappearance of the stretching frequency at 1700-1708 cm⁻¹ assigned to $\nu(\text{COOH})$ and appearance of new bands of the $(-\text{CO}_2)$ group at 1590 and 1385 cm⁻¹, is consistent with carboxylate coordination with the metal atom. The shifting of medium to high intensity bands of enolic C-O bond towards higher frequency (1339-1368 cm⁻¹) and appearance of new bands at 510-540 cm⁻¹ support the formation of M-O bond via de-protonation.[22] The presence of coordinated water molecule in the complex is indicated by the appearance of a broad band at 3226-3460 cm⁻¹ and two weak bands in the region 754-784 cm⁻¹ and 700-718 cm⁻¹ due to $(-\text{OH})$ rocking and wagging mode of vibrations, respectively. Apart from this, the infrared spectra of the VO²⁺ also exhibit a strong band at 996-970 cm⁻¹ which is

attributed to the terminal V=O bond. These overall data suggest that the azomethine-N, carboxylate-O and enolic-O groups are involved in coordination with the metal(II) ion in complexes (C₁-C₄).

Table 1. The important infrared frequencies (in cm⁻¹) of oxovanadium complexes.

Code	Complex	$\nu(\text{C}=\text{N})$	$\nu(\text{M}-\text{N})$	$\nu(\text{M}-\text{O})$	$\nu(\text{C}-\text{O})$	$\nu(\text{C}=\text{C})$
C ₁	[VO(L ₁)(OH ₂)]	1590s	455m	510m	1232m	1624
C ₂	[VO(L ₂)(OH ₂)]	1594m	462m	540m	1103m	1630
C ₃	[VO(L ₃)(OH ₂)]	1591m	458m	535m	1101m	1623
C ₄	[VO(L ₄)(OH ₂)]	1594m	460m	512m	1093m	1629

3.2. ¹H NMR Spectra

The ¹H NMR spectral data along with the possible assignments are reported in "Experimental." All the protons were found as to be in their expected region.[23] The ¹H-NMR spectra of Schiff bases exhibit singlet signals at 12.2-12.8, 11.2-11.3 and 2.1 ppm due to enolic OH, COOH, and methyl group attached with azomethine carbon atom, protons respectively. The resonance due to phenolic -OH in L₁ was observed at 8.7 ppm. The signal due to proton attached with alkene carbon was observed at 5.1-5.2 ppm. In L₃ a multiplet signal was found at 5.3 ppm due to primary amine. A multiplet signal at 3.4 ppm was observed due to proton of secondary amine in L₄. The ¹H NMR of VO(IV) complexes was not recorded as all the complexes were paramagnetic in nature.[24] The conclusions drawn from these studies lend further support to the mode of bonding discussed in their IR spectra. The number of protons calculated from the integration curves and those obtained from the values of the expected CHN analyses agree with each other.

3.3. ¹³C NMR Spectra

¹³C NMR spectra of amino-acid derived Schiff bases were recorded in DMSO and the data are given in the Experimental. The signal due to the azomethine carbon atom appears at δ 162.1 and 168.2 ppm in all the prepared compounds which confirms the condensation reaction between acetylacetone and amino-acid. Additional signal in the ¹³C-NMR spectra of the Schiff bases appears at 172.2-178.3 and 80.1-85.2 ppm indicating the alkene bond.

3.4. Thermogravimetric Analysis

The TGA curves of the complexes were recorded in the temperature range 30-1000°C and are given in Table 2. The dehydration of the water molecules found outside the coordination sphere usually take place in the temperature range 25-220 °C. Coordinated waters are usually eliminated at higher temperatures (100-315 °C).[25] The organic part of the complex may decompose in one or more steps with

the possibility of the formation of one or two Intermediates, calculated and estimated mass losses are comparable. The thermal decomposition of [VO(L)H₂O] occurs in two steps. In the first step, TGA curve of oxovanadium complexes shows loss of a water molecule in the temperature range of 145-280 °C which corresponds to a weight loss of 5.20-6.33%. In the second step a continuous weight loss in the temperature range 280-595 °C is observed due to loss of Schiff base ligand. The final residue with attaining a constant weight roughly corresponds to V₂O₅.

3.5. Conductance

The molar conductivity measurements were carried out in DMSO at 10⁻³ molar concentration. The Molar conductance values for oxovanadium(IV) complexes are gives in Table 3. All the complexes (C₁-C₄) have molar conductance in the range 10-25 $\Omega^{-1} \text{ cm}^2 \text{ mol}^{-1}$ indicating non-electrolytic nature of the complexes.[26]

Table 2. Thermogravimetric analysis (TGA) results of oxovanadium complexes.

Code	Temperature range °C	Mass loss % Found (Calcd.)	Assignment
C ₁	145-280	5.23 (5.20)	Loss of H ₂ O
	284-575	75.05 (74.98)	Loss L ₁
C ₂	145-280	6.35 (6.33)	Loss of H ₂ O
	285-584	70.02 (69.22)	Loss L ₂
C ₃	145-280	5.86 (5.78)	Loss of H ₂ O
	282-595	72.15 (72.02)	Loss L ₃
C ₄	145-280	5.36 (5.30)	Loss of H ₂ O
	284-575	74.49 (74.44)	Loss L ₄

3.6. Magnetic Moments and Electronic Spectra

The magnetic moment values of oxovanadium(IV) complexes were carried out at room temperature and are given in Table 4. The magnetic susceptibilities of VO(IV) complexes were found to be in the range of 1.74-1.86, suggesting the paramagnetic nature of the complexes. These magnetic susceptibilities of the complexes are consistent with square-pyramidal geometry around the central metal ion.[27] Electronic absorption spectral data of oxovanadium(IV) complexes along with their assignment are given in the Table IV. The electronic spectra of VO(IV) complexes exhibit three low intensities bands in the ranges 11370-13590 cm⁻¹, 15900-17150 cm⁻¹ and 22400-23900 cm⁻¹ which can be assigned ²b_{2g}→²e_g, ²b_{2g}→²b_{1g} and ²b_{2g}→²a_{1g} transitions respectively, characteristic of a square pyramidal ligand field around VO(IV) ion. The electronic spectra also exhibit a fourth high energy band around 29200-30250 cm⁻¹, which can be assigned as a LMCT transition which is consistent with square pyramidal geometry around the V⁴⁺.

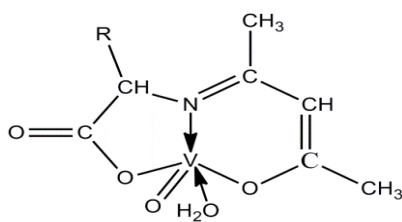


Figure 5. Proposed structure of oxovanadium(IV) complexes

Table 3. Physical and Analytical data of oxovanadium complexes.

Code	Molar conductance ($\text{Ohm}^{-1} \text{cm}^2 \text{mol}^{-1}$)	Mag. moments (μ_{eff} in BM)	Calcd. (Found) %	Calcd. (Found) %	Calcd. (Found) %
			C	H	N
C ₁	13.1	1.74	48.57 (48.41)	4.95 (5.01)	4.05 (4.12)
C ₂	14.2	1.86	38.04 (38.11)	5.32 (5.36)	4.93 (4.99)
C ₃	10.0	1.82	42.45 (42.41)	6.48 (6.52)	9.00 (9.06)
C ₄	25.6	1.83	38.94 (38.96)	5.94 (6.01)	16.52 (16.59)

Table 4. Electronic spectral data along with possible assignments of oxovanadium(IV) complexes.

Code	$\lambda_{\text{max}} \text{cm}^{-1}$	Assignment	Code	$\lambda_{\text{max}} \text{cm}^{-1}$	Assignment
C ₁	11540-13460	${}^2b_{2g} \rightarrow {}^2e_g$	C ₃	11520-13360	${}^2b_{2g} \rightarrow {}^2e_g$
	16200-16900	${}^2b_{2g} \rightarrow {}^2b_{1g}$		15900-17150	${}^2b_{2g} \rightarrow {}^2b_{1g}$
	22400-23900	${}^2b_{2g} \rightarrow {}^2a_{1g}$		22640-23780	${}^2b_{2g} \rightarrow {}^2a_{1g}$
	29400-29980	LMCT		29350-30100	LMCT
C ₂	11370-13590	${}^2b_{2g} \rightarrow {}^2e_g$	C ₄	11390-13250	${}^2b_{2g} \rightarrow {}^2e_g$
	15990-17000	${}^2b_{2g} \rightarrow {}^2b_{1g}$		15990-16970	${}^2b_{2g} \rightarrow {}^2b_{1g}$
	22780-23500	${}^2b_{2g} \rightarrow {}^2a_{1g}$		22600-23750	${}^2b_{2g} \rightarrow {}^2a_{1g}$
	29200-30250	LMCT		29340-30240	LMCT

3.7. Antibacterial and Antifungal Activities

The antibacterial and antifungal activity results of the Schiff bases (L₁-L₄) and their oxovanadium(IV) complexes (C₁-C₄) are given in Fig. 6 and 7. Antimicrobial results showed that all the synthesized compounds possess biological activity. The activity generally increases with increasing the concentration of the compounds. The data revealed that the activity of the ligand enhanced on complexation but less than the standard used. This enhancement in the activity is rationalized on the basis of the structures of, (L₁-L₄) by possessing an additional azomethine (-C=N) linkage which imports in elucidating the mechanism of transamination and resamination reactions in biological system. It has also known that the

ligands with nitrogen and oxygen donor systems might inhibit enzyme production, since the enzymes which require these groups for their activity appear to be especially more susceptible to deactivation by the metal ions upon chelation.[28] On chelation the polarity of the metal ion will be reduced to a greater extent due to overlap of the ligand orbital and partial sharing of positive charges of metal ion with donor groups. Further it increases the delocalisation of pi electrons over the whole chelate ring and enhances the lipophilicity of the complex. This again enhances the penetration of complexes into lipid membrane and blocking the metal bonding sites on enzymes of micro organisms, hence increases the biological activity.[29]

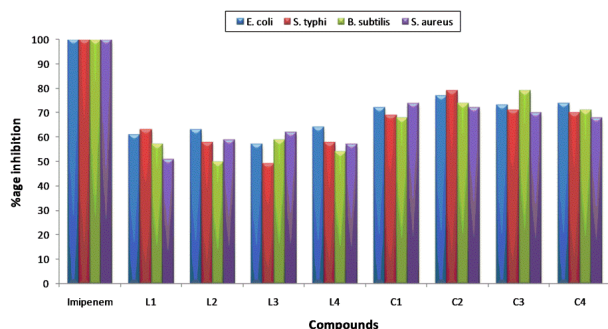


Figure 6. In vitro antibacterial spectrum of Schiff bases (L_1 - L_4), oxovanadium(IV) complexes and Imipenem (Std.) at $100 \mu\text{gml}^{-1}$ concentration.

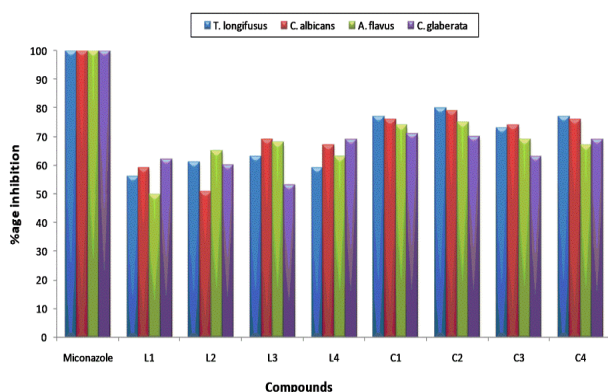


Figure 7. In vitro antifungal spectrum of Schiff bases (L_1 - L_4), oxovanadium(IV) complexes and Miconazole (Std.) at $100 \mu\text{gml}^{-1}$ concentration.

3.8. Cytotoxicity

All the synthesized compounds were screened for their cytotoxicity (brine shrimp bioassay) using the protocol of Meyer et al. The cytotoxicity results are given in Table 5. In case of cytotoxic activity it was observed that all the compounds and their oxovanadium(IV) complexes gave values of LD_{50} therefore can be considered as non-cytotoxic.

Table 5. Brine shrimp bioassay data of the Schiff bases (L_1 - L_4) and their oxovanadium complexes (C_1 - C_4).

Code	LD_{50} (M)	Code	LD_{50} (M)
L_1	$>3.36 \times 10^{-3}$	C_1	$>2.61 \times 10^{-3}$
L_2	$>2.03 \times 10^{-3}$	C_2	$>2.02 \times 10^{-3}$
L_3	$>2.94 \times 10^{-3}$	C_3	$>1.30 \times 10^{-3}$
L_4	$>2.87 \times 10^{-3}$	C_4	$>3.11 \times 10^{-3}$

4. Conclusion

The synthesized Schiff bases act as tri-identate ligands. The IR, TGA, conductivity, magnetic and electronic studies confirm that the vanadyl ion is coordinated to azomethine-N, deprotonated-O of carboxylic group and enolic-O group of

acetylacetone. All the derivatives and their oxovanadium(IV) complexes were evaluated *in vitro* against four bacterial (two Gram-negative, two Gram-positive) and four fungal strains. All the compounds and their metal complexes showed significant antibacterial and antifungal potency. Brine shrimp bioassay data revealed that all the compounds were non-cytotoxic against *Artemia salina*.

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